

eISBN: 978-1-68108-289-9
ISBN: 978-1-68108-290-5

eISSN: 2215-0803
ISSN: 2451-8905

Frontiers in Clinical Drug Research (Anti-Cancer Agents) ^{Volume 3}



Editor:
Atta-ur-Rahman, FRS

Bentham  Books

**Frontiers in Clinical Drug
Research - Anti-Cancer Agents**
Volume 3

Edited By

Atta-ur-Rahman, *FRS*

Kings College

University of Cambridge

Cambridge

UK

Frontiers in Clinical Drug Research - Anti-Cancer Agents

[Volume 3]

ISSN (Online): 2215-0803

ISSN (Print): 2451-8905

Frontiers in Clinical Drug Research - Anti-Cancer Agents

Editor: Atta-ur-Rahman, *FRS*

ISBN (eBook): 978-1-68108-289-9

ISBN (Print): 978-1-68108-290-5

©[2016], Bentham eBooks imprint.

Published by Bentham Science Publishers – Sharjah, UAE. All Rights Reserved.

BENTHAM SCIENCE PUBLISHERS LTD.

End User License Agreement (for non-institutional, personal use)

This is an agreement between you and Bentham Science Publishers Ltd. Please read this License Agreement carefully before using the ebook/echapter/ejournal (“**Work**”). Your use of the Work constitutes your agreement to the terms and conditions set forth in this License Agreement. If you do not agree to these terms and conditions then you should not use the Work.

Bentham Science Publishers agrees to grant you a non-exclusive, non-transferable limited license to use the Work subject to and in accordance with the following terms and conditions. This License Agreement is for non-library, personal use only. For a library / institutional / multi user license in respect of the Work, please contact: permission@benthamscience.org.

Usage Rules:

1. All rights reserved: The Work is the subject of copyright and Bentham Science Publishers either owns the Work (and the copyright in it) or is licensed to distribute the Work. You shall not copy, reproduce, modify, remove, delete, augment, add to, publish, transmit, sell, resell, create derivative works from, or in any way exploit the Work or make the Work available for others to do any of the same, in any form or by any means, in whole or in part, in each case without the prior written permission of Bentham Science Publishers, unless stated otherwise in this License Agreement.
2. You may download a copy of the Work on one occasion to one personal computer (including tablet, laptop, desktop, or other such devices). You may make one back-up copy of the Work to avoid losing it. The following DRM (Digital Rights Management) policy may also be applicable to the Work at Bentham Science Publishers’ election, acting in its sole discretion:
 - 25 ‘copy’ commands can be executed every 7 days in respect of the Work. The text selected for copying cannot extend to more than a single page. Each time a text ‘copy’ command is executed, irrespective of whether the text selection is made from within one page or from separate pages, it will be considered as a separate / individual ‘copy’ command.
 - 25 pages only from the Work can be printed every 7 days.
3. The unauthorised use or distribution of copyrighted or other proprietary content is illegal and could subject you to liability for substantial money damages. You will be liable for any damage resulting from your misuse of the Work or any violation of this License Agreement, including any infringement by you of copyrights or proprietary rights.

Disclaimer:

Bentham Science Publishers does not guarantee that the information in the Work is error-free, or warrant that it will meet your requirements or that access to the Work will be uninterrupted or error-free. The Work is provided "as is" without warranty of any kind, either express or implied or statutory, including, without limitation, implied warranties of merchantability and fitness for a particular purpose. The entire risk as to the results and performance of the Work is assumed by you. No responsibility is assumed by Bentham Science Publishers, its staff, editors and/or authors for any injury and/or damage to persons or property as a matter of products liability, negligence or otherwise, or from any use or operation of any methods, products instruction,

advertisements or ideas contained in the Work.

Limitation of Liability:

In no event will Bentham Science Publishers, its staff, editors and/or authors, be liable for any damages, including, without limitation, special, incidental and/or consequential damages and/or damages for lost data and/or profits arising out of (whether directly or indirectly) the use or inability to use the Work. The entire liability of Bentham Science Publishers shall be limited to the amount actually paid by you for the Work.

General:

1. Any dispute or claim arising out of or in connection with this License Agreement or the Work (including non-contractual disputes or claims) will be governed by and construed in accordance with the laws of the U.A.E. as applied in the Emirate of Dubai. Each party agrees that the courts of the Emirate of Dubai shall have exclusive jurisdiction to settle any dispute or claim arising out of or in connection with this License Agreement or the Work (including non-contractual disputes or claims).
2. Your rights under this License Agreement will automatically terminate without notice and without the need for a court order if at any point you breach any terms of this License Agreement. In no event will any delay or failure by Bentham Science Publishers in enforcing your compliance with this License Agreement constitute a waiver of any of its rights.
3. You acknowledge that you have read this License Agreement, and agree to be bound by its terms and conditions. To the extent that any other terms and conditions presented on any website of Bentham Science Publishers conflict with, or are inconsistent with, the terms and conditions set out in this License Agreement, you acknowledge that the terms and conditions set out in this License Agreement shall prevail.

Bentham Science Publishers Ltd.

Executive Suite Y - 2

PO Box 7917, Saif Zone

Sharjah, U.A.E.

Email: subscriptions@benthamscience.org



CONTENTS

PREFACE	i
LIST OF CONTRIBUTORS	iii
CHAPTER 1 TUMOR RESISTANCE MECHANISMS TO INHIBITORS TARGETING THE EPIDERMAL GROWTH FACTOR RECEPTOR– PART I: EXTRACELLULAR MOLECULES	3
<i>Rodney B. Luwor</i>	
1. INTRODUCTION	4
2. LINKING THE EGFR WITH CANCER THERAPUTICS	7
3. THERAPEUTIC AGENTS TARGETING THE EGFR	10
3.1. Cetuximab/Erbitux (Bristol-Myers Squibb and Eli Lilly and Company)	10
3.2. Panitumumab/Vectibix (Amgen Inc)	12
3.3. Lapatinib/Tykerb (GlaxoSmithKline)	13
3.4. Gefitinib/Iressa (AstraZeneca)	13
3.5. Erlotinib/Tarceva (Genentech)	15
3.6. Afatinib/Gilotrif (Boehringer Ingelheim Pharmaceuticals)	18
4. BIOMARKERS PREDICTING SENSITIVITY AND RESISTANCE TO ANTI-EGFR THERAPIES - LIGANDS OF THE EGFR AS PREDICTORS OF RESPONSE TO ANTI-EGFR THERAPY	19
4.1. Amphiregulin (AREG) and Epregrulin (EREG)	20
4.2. Transforming Growth Factor Alpha (TGF α)	23
4.3. Epidermal Growth Factor (EGF)	24
4.4. Heparin-Binding EGF-LIKE Growth Factor (HB-EGF)	26
4.5. Betacellulin (BTC) and Epigen (EPGN)	27
5. BIOMARKERS PREDICTING SENSITIVITY AND RESISTANCE TO ANTI-EGFR THERAPIES - ALTERATIONS IN ERBB RECEPTORS FAMILY MEMBERS	28
5.1. Epidermal Growth Factor Receptor (EGFR)	29
5.1.1. EGFR Gene and Protein Expression	29
5.1.2. EGFR Gene Copy Number (GCN)	35
5.1.3. EGFR Mutations	39
5.2. HER2 (ErbB2)	56
5.3. HER3, HER4 and Neuregulins (NRG)	59
6. ALTERATIONS IN NON-ERBB RECEPTOR FAMILY MEMBERS	63
6.1. Hepatocyte Growth Factor/Scatter Factor (HGF)/MET	64
6.2. Insulin-like Growth Factor (IGF)/IGF-R	68
6.3. Fibroblast Growth Factor (FGF)/FGFR	72
6.4. Platelet-derived Growth Factor (PDGF)/PDGFR	73
6.5. Vascular Endothelial Growth Factor (VEGF)/VEGFR	74
6.6. GAS6/AXL	77
CONCLUDING REMARKS	80
CONFLICT OF INTEREST	81
ACKNOWLEDGEMENTS	81
ABBREVIATIONS	81
REFERENCES	82
CHAPTER 2 TUMOR RESISTANCE MECHANISMS TO INHIBITORS TARGETING THE EPIDERMAL GROWTH FACTOR RECEPTOR – PART II: INTRACELLULAR MOLECULES	142
<i>Rodney B. Luwor</i>	
1. INTRODUCTION	143
2. INTRACELLULAR TRAFFICKING OF THE EGFR AND RESISTANCE TO ANTI-EGFR THERAPY	146

3. THE RAS/REF/MEK/ERK SIGNALING PATHWAY AND RESISTANCE TO ANTI-EGFR THERAPY	150
3.1. Growth Factor Receptor-bound Protein 2 (Grb2)	150
3.2. RAS	151
3.2.1. <i>The Role of K-RAS in Primary Resistance</i>	151
3.2.2. <i>The Role of K-RAS in Acquired Resistance</i>	159
3.3. RAF	160
3.4. MAPK Extracellular Signal-regulated Kinase (MEK) and Extracellular Signal-regulated Kinase (ERK1/2)	164
3.5. Dual-Specificity MAP Kinase Phosphatases (DUSPs)	166
4. THE PI3-K SIGNALING PATHWAY AND RESISTANCE TO ANTI-EGFR THERAPY	166
4.1. Phosphatidylinositol 3-kinase (PI3-K)	167
4.2. PTEN	175
5. ANALYSIS OF SEVERAL BIOMARKERS IN COMBINATION AND RESISTANCE TO ANTI-EGFR THERAPY	181
6. THE JAK/STAT SIGNALING PATHWAY AND RESISTANCE TO ANTI-EGFR THERAPY	183
CONCLUDING REMARKS	186
CONFLICT OF INTEREST	187
ACKNOWLEDGEMENTS	187
ABBREVIATIONS	188
REFERENCES	189

CHAPTER 3 CHEMOTHERAPEUTIC, IMMUNOLOGIC, AND MOLECULARLY TARGETED THERAPY FOR THE TREATMENT OF ADVANCED MELANOMA 217

<i>Said C. Azoury, David M. Straughan, Robert D. Bennett and Vivek Shukla</i>	
INTRODUCTION	218
CHEMOTHERAPEUTIC AGENTS	220
Alkylating Agents	220
<i>Dacarbazine</i>	220
<i>Temozolomide</i>	221
Platinum Compounds	222
Taxanes	222
Nitrosureas	223
Chemotherapy in Combination	223
IMMUNOTHERAPY	225
Interferon Alpha	226
Interleukin-2 (IL-2)	227
Vaccine Therapy	230
<i>Peptide Vaccines</i>	230
<i>Adoptive Cell Therapy</i>	238
<i>Immune Checkpoint Inhibitors</i>	239
<i>Immune Checkpoint Inhibitors in Combination</i>	244
MOLECULARLY TARGETED THERAPY	246
<i>MAPK Signaling Pathway</i>	246
<i>BRAF Inhibitors</i>	248
<i>MEK Inhibitors</i>	250
<i>C-KIT Inhibitors</i>	251
<i>Molecularly Targeted Therapies in Combination</i>	251
CONCLUSION	253
CONFLICT OF INTEREST	254
ACKNOWLEDGEMENTS	254
REFERENCES	254

CHAPTER 4 TARGETING THE WARBURG EFFECT FOR CANCER THERAPY: A LONG AND WINDING ROAD 271

Patricia L. Abreu and Ana M. Urbano

1. THERAPEUTICAL APPROACHES TO CANCER: FROM CHEMOTHERAPY TO TARGETED THERAPIES	272
2. BIOENERGETICS: A VERY BRIEF OVERVIEW	277
3. THE WARBURG EFFECT: CURRENT PERSPECTIVE	282
4. CURRENT AND PAST EFFORTS AT TARGETING THE WARBURG EFFECT FOR CANCER THERAPY	288
CONCLUDING REMARKS	310
NOTES	312
CONFLICT OF INTEREST	312
ACKNOWLEDGEMENTS	312
ABBREVIATIONS	312
REFERENCES	313

CHAPTER 5 IMMUNOTHERAPY STRATEGIES IN FOLLICULAR LYMPHOMA: ANTIBODIES, VACCINES AND CELLS 325

Nicolás Martínez-Calle, Ascensión López Díaz de Cerio, Susana Inogés, Esther Pena, Ricardo García-Muñoz and Carlos Panizo

1. INTRODUCTION	326
2. BIOLOGY OF FOLLICULAR LYMPHOMA	327
2.1. Histopathology of FL	327
2.2. Follicular Lymphoma-cell Origin	327
2.3. Follicular Lymphoma Microenvironment	328
3. PREVIOUS CONSIDERATIONS BEFORE THERAPY	331
3.1. Who should be Treated, and When?	331
3.2. How to define Treatment Intensity	333
4. FIRST LINE TREATMENT	334
4.1. Early-stage Disease	334
4.2. Induction Treatment for Advanced Stages	335
4.3. Consolidation Treatment	338
4.3.1. Interferon Maintenance	338
4.3.2. Rituximab Maintenance	338
4.3.3. Autologous Stem-cell Transplantation	340
4.3.4. Radioimmunotherapy	340
5. RELAPSE TREATMENT	342
5.1. Choice of Rescue Treatment	342
5.2. Bortezomib for FL Relapse	342
5.3. Autologous Stem-cell Transplantation for FL Relapse	343
5.4. Radioimmunotherapy for FL Relapse	344
5.5. Rituximab Maintenance for FL Relapse	344
5.6. Role of Allogeneic SCT	345
6. NOVEL AND EMERGING PHARMACOLOGICAL AGENTS	345
7. INNOVATIVE IMMUNOTHERAPY STRATEGIES FOR TREATMENT OF FL	347
7.1. Passive Immunotherapy: Adoptive Cell Therapy	348
7.1.1. LAK Cells	348
7.1.2. NK Cells	349
7.1.3. Antigen-specific T Cells	350
7.1.4. Treatment Combinations: Rituximab and Effector Cells	352
7.2. Active Immunotherapy Strategies: Vaccines	353
7.2.1. Idiotypic Protein Vaccines	354
7.2.2. DNA Vaccines	355

7.2.3. Dendritic Cell Vaccines	356
7.2.4. Membrane Proteoliposomal Vaccines	357
7.2.5. Strategies to Enhance Immunogenicity	357
7.3. New Targets for Immunotherapy	358
7.4. Future Directions in Vaccine Development	359
CONCLUDING REMARKS	360
CONFLICT OF INTEREST	360
ACKNOWLEDGEMENTS	360
REFERENCES	360

CHAPTER 6 EPIGENETICS AND CANCER CELL METABOLISM: CROSS-TALK AND THERAPEUTIC OPPORTUNITIES 386

Chi Chun Wong and Jun Yu

INTRODUCTION	387
CANCER EPIGENETICS AND ROLE IN CARCINOGENESIS	389
DNA Methylation	389
Histone Modifications	390
Non-coding RNA	392
CANCER METABOLISM	393
The Warburg's Hypothesis: Deregulated Metabolism in Cancer	393
Pathways Leading to Altered Metabolism in Tumors	395
Epigenetics and Metabolism: Crosstalk	398
MODULATION OF METABOLISM BY EPIGENETIC DYS-REGULATION	399
Modulation of Metabolism by DNA Methylation	399
Modulation of Metabolism by Histone Modifications	402
Modulation of Metabolism by miRNAs	404
EPIGENETIC REGULATION BY THE AVAILABILITY OF CO-SUBSTRATES	410
Acetyl-CoA and Histone Acetylation	410
S-adenosylmethionine and DNA and Histone Methylation	412
EPIGENETIC DYSREGULATION INDUCED BY ONCOMETABOLITES	416
2-Hydroxylglutarate Production in IDH1 and IDH2 Mutant Cancers	416
2-Hydroxylglutarate Modulates DNA Methylation	418
2-Hydroxylglutarate Modulates Histone Methylation	419
Succinate and Fumarate Modulate DNA and Histone Methylation	421
THERAPEUTIC OPPORTUNITIES	423
DNMT Inhibitors	423
HDAC Inhibitors (HDACi)	424
miRNA Targeting Agents	424
IDH1 and IDH2 Inhibitors	425
CONCLUDING REMARKS	426
CONFLICT OF INTEREST	428
ACKNOWLEDGEMENTS	428
REFERENCES	428

CHAPTER 7 APOPTOSIS TARGETING THERAPEUTICS IN CLINICAL TRIALS 454

N.S. Hari Narayana Moorthy, C. Karthikeyan and Piyush Trivedi

CANCER	455
APOPTOSIS	456
MECHANISMS OF APOPTOSIS	456
TARGETING P53	458
p53-based Gene Therapy	459
p53-based Immunotherapy	460
p53-based Drug Therapy	460

ANTI-APOPTOTIC PROTEINS	460
Targeting the MDM2-p53 Complex	460
<i>Nutlins</i>	460
<i>RG7112</i>	463
<i>MK-8242 and MI-888</i>	463
<i>MI-219</i>	463
<i>SAR405838 (MI-77301)</i>	464
<i>CGM097</i>	465
<i>Other MDM2 Inhibitors</i>	465
Targeting the IAPs	466
<i>OGX-011</i>	467
<i>GDC-0152 and GDC-0917 (CUDC-427) (Genentech)</i>	468
<i>AE40826/HGS1029 (Aegera/Human Genome Sciences)</i>	470
<i>LCL161 (Novartis)</i>	470
<i>SM-406/AT-406/Debio 1143 (University of Michigan/Ascenta/DebioPharm)</i>	471
<i>SM-164</i>	471
<i>LBW242</i>	471
<i>Other IAP Antagonists</i>	472
Targeting the BCL Family of Proteins	472
<i>ABT-737 and ABT-263 (navitoclax)</i>	474
<i>ABT-199 (GDC-0199/venetoclax, AbbVie)</i>	475
<i>Obatoclax Mesylate (GX15-070) (Gemin X/Teva)</i>	476
<i>AT-101 R-(-)-gossypol (Ascenta)</i>	477
OTHER APOPTOSIS INDUCERS	478
CONFLICT OF INTEREST	480
ACKNOWLEDGEMENTS	480
REFERENCES	480
SUBJECT INDEX	490

PREFACE

The third volume of *Frontiers in Clinical Drug Research - Anti-Cancer Agents* presents seven cutting edge reviews on recent developments in various therapeutic approaches against different types of cancer.

Studies have revealed that the Epidermal Growth Factor Receptor (EGFR) is involved in the pathogenesis and progression of different types of carcinoma. Tumor resistance to agents targeting the Epidermal Growth Factor Receptor (EGFR) is common, and is well recognized as a major challenge. In first two consecutive chapters, Rodney B. Luwor provides an overview of the progress in targeting the EGFR that will lead to overall refractory outcomes to anti-EGFR therapies. In Chapter 1 he discusses on the resistance mechanisms driven by alterations in ligand and receptors of the EGFR family as well as on the cross-talk between EGFR receptors and non-EGFR family members. In Chapter 2 the same author describes the current understanding regarding the resistance mechanisms mediated by alterations in substrates downstream of the EGFR. Luwor has also reviewed the other intracellular mechanisms that mediate both sensitivity and resistance outcomes to anti-EGFR agents in this chapter.

Melanoma is the most dangerous form of skin cancer that develops when unrepaired DNA damage to skin cells triggers mutations, which lead to the formation malignant tumors. In Chapter 3 Shukla *et al.*, present a comprehensive review on the chemotherapeutic, immunologic, and molecularly targeted therapy approaches to the treatment of advanced melanoma.

In various tumor cells, there is increased aerobic glycolysis that represents a major biochemical alteration associated with malignant transformation. This phenomenon is known as the Warburg effect. 18F-deoxyglucose positron emission tomography (18FDG–PET), a metabolic imaging technique, is based on the avidity of cancer cells for glucose; currently, it represents the only successful exploitation of the Warburg effect for medical purposes. In Chapter 4, Abreu and Urbano focus on past and current efforts to target the Warburg effect for selective anti-cancer therapeutics.

Follicular lymphoma (FL) is a B-cell lymphoma and the most common slow-growing form of non-Hodgkin lymphoma (NHL). Studies suggest that immunotherapy, radioimmunotherapy and vaccines result in high response rates and survival in FL patients. Chapter 5 by Panizo *et al.*, briefly describes the biology and conventional treatment of follicular lymphoma with immunochemotherapy. They also discuss novel immunotherapy strategies (active and passive) for the treatment of follicular lymphoma.

ii

The progression of cancer involves epigenetic abnormalities along with genetic alterations. The manipulation of epigenetic alterations holds great promise for the prevention, detection, and therapy of cancer. Evidence indicates that the activities of key epigenetic regulators including DNA methyltransferases and histone modification enzymes are sensitive to cellular metabolism. Wong and Yu in Chapter 6 discuss that the cross-talk between epigenetics and cancer cell metabolism may reveal novel therapeutic opportunities. They also highlight their implications in oncogenesis, and potential therapeutic approaches to target these cancer specific abnormalities.

Apoptosis is a programmed cell death, which involves various biochemical events that lead to characteristic cell changes and death. Dysfunctions of apoptosis pathways promote oncogenesis as well as confer resistance of cancer cells to most conventional therapies. In Chapter 7 by Moorthy *et al.* focus their discussion small molecular anticancer drugs, especially target proteins, responsible for apoptosis.

I hope that the current volume of this book series will provide fresh insights into development of new recent approaches to anti-cancer therapy for interested researchers and pharmaceutical scientists. I would like to thank the editorial staff, particularly Mr. Mahmood Alam (Director Publications) and Mr. Shehzad Naqvi (Senior Manager Publications) for their hard work and dedicated efforts.

Atta-ur-Rahman, FRS
Kings College
University of Cambridge
Cambridge
UK

List of Contributors

- Ana M. Urbano** Unidade de Química-Física Molecular and Departamento de Ciências da Vida, Faculdade de Ciências e Tecnologia, Universidade de Coimbra, Coimbra, Portugal
- Ascensión López Díaz de Cerio** Hematology Department, Clínica Universidad de Navarra, Pamplona, Spain
- Carlos Panizo** Hematology Department, Clínica Universidad de Navarra, Pamplona, Spain
- Chi Chun Wong** Department of Medicine and Therapeutics, State Key Laboratory of Digestive Disease, Li Ka Shing Institute of Health Sciences, Shenzhen Research Institute, The Chinese University of Hong Kong, Hong Kong
- C. Karthikeyan** Departamento de Química e Bioquímica, Faculdade de Ciências, Universidade do Porto, s/n, Rua do Campo Alegre, 4169-007 Porto, Portugal
- David M. Straughan** Department of Surgery, University of South Florida, Morsani College of Medicine, Tampa, FL, USA
- Esther Pena** Hematology Department, Complejo Hospitalario de Navarra, Pamplona, Spain
- Jun Yu** Department of Medicine and Therapeutics, State Key Laboratory of Digestive Disease, Li Ka Shing Institute of Health Sciences, Shenzhen Research Institute, The Chinese University of Hong Kong, Hong Kong
- N.S. Hari Narayana Moorthy** Departamento de Química e Bioquímica, Faculdade de Ciências, Universidade do Porto, s/n, Rua do Campo Alegre, Porto, Portugal; School of Pharmaceutical Sciences, Rajiv Gandhi Proudhyogiki Vishwavidyalaya, Airport Bypass Road, Gandhi Nagar, Bhopal, India
- Nicolás Martínez-Calle** Hematology Department, Clínica Universidad de Navarra, Pamplona, Spain
- Patrícia L. Abreu** Unidade de Química-Física Molecular and Departamento de Ciências da Vida, Faculdade de Ciências e Tecnologia, Universidade de Coimbra, Coimbra, Portugal
- Piyush Trivedi** School of Pharmaceutical Sciences, Rajiv Gandhi Proudhyogiki Vishwavidyalaya, Bhopal (MP)-462033, India
- Ricardo García-Muñoz** Hematology Department, Hospital San Pedro, Logroño, Spain
- Robert D. Bennett** Department of Surgery, University of South Florida, Morsani College of Medicine, Tampa, FL, USA
- Rodney B. Luwor** Department of Surgery, The Royal Melbourne Hospital, The University of Melbourne, Parkville, Victoria 3050, Australia
- Saïd C. Azoury** Department of Surgery, The Johns Hopkins Hospital, Baltimore, MD, USA

iv

Susana Inogés

Hematology Department, Clínica Universidad de Navarra, Pamplona, Spain

Vivek Shukla

Thoracic and GI Oncology Branch, National Cancer Institute, National Institutes of Health, Bethesda, MD, USA

Tumor Resistance Mechanisms to Inhibitors Targeting the Epidermal Growth Factor Receptor— Part I: Extracellular Molecules

Rodney B. Luwor*

Department of Surgery, The Royal Melbourne Hospital, The University of Melbourne, Parkville, Victoria 3050, Australia

Abstract: Since its discovery several decades ago, the Epidermal Growth Factor Receptor (EGFR) has become one of the most extensively studied receptor tyrosine kinases. However, despite continued insight into the cancer promoting properties of the EGFR and its downstream signalling substrates, clinical use of agents targeting the EGFR continue to yield modest outcomes. Clinically, approved anti-EGFR therapeutics can successfully inhibit receptor activation. However major tumour regression is observed in only 10-30% of advanced unselected cancer patients, with most patients showing no therapeutic benefit. Furthermore, those who initially respond commonly relapse presenting with reoccurrence of tumours that are frequently resistant to the original therapy. In addition, the standard course of treatment of such agents is estimated to cost between “US \$15,000-80,000/patient” for an improved overall survival of only 1-2 months. Therefore, it is both medically and financially critical to determine the true molecular mechanisms of tumour resistance, and how it can be overcome. In these 2 back-to-back chapters, we will provide an overview of the progress made in targeting the EGFR and discuss the challenges presented by the numerous molecular mechanisms currently identified, leading to overall refractory outcomes to anti-EGFR therapeutics. In this chapter (Part I) we will specifically focus on the resistance mechanisms driven by alterations in ligand and receptors of the EGFR family and cross-talk between EGFR receptors and non-EGFR family members.

Keywords: Afatinib, Cancer, Cetuximab, Epidermal Growth Factor Receptor,

* **Corresponding author Rodney B. Luwor:** Department of Surgery, The Royal Melbourne Hospital, The University of Melbourne, Parkville, Victoria 3050, Australia; Tel: +613 8344 3027 Fax: +613 9347 6488; E-mail: rluwor@unimelb.edu.au

Erlotinib, Gefitinib, Lapatinib, Panitumumab, Resistance, Signaling, Therapeutics, Tumor.

1. INTRODUCTION

Since the discovery of the Epidermal Growth Factor (EGF) in 1962 by Stanley Cohen and colleagues [1] tremendous advances in our understanding of the sophisticated interactions between growth factors and their accompanying cell surface receptors have been made. One of the most intensely studied classes of receptors is the HER or ErbB family [2]. This family consists of four members, the Epidermal Growth Factor Receptor (EGFR) (also referred to as ErbB1 or HER1) [3], HER2 (p185^{Neu} or ErbB2) [4], HER3 (ErbB3) [5] and HER4 (ErbB4) [6]. All 4 family members share a similar overall structure consisting of an extracellular domain with 2 cysteine-rich regions, a single membrane-spanning region and a cytoplasmic domain containing multiple tyrosine residues that are phosphorylated upon receptor activation [7, 8].

The *EGFR* gene is located on the short arm of chromosome 7 [9, 10], and encodes an 1186 amino acid long, 140 KDa polypeptide chain [3, 11], which contains approximately 30 – 40 KDa of N-linked oligosaccharides [12, 13]. A single 23 amino acid long hydrophobic sequence transverses the cell membrane. The extracellular N-terminal end (amino acids 1 - 621) can be divided into four domains (I-IV) [14, 15]. The intracellular C-terminal region (amino acids 645 - 1186) is responsible for tyrosine kinase activity and regulatory functions [16].

Currently eight ligands have been identified to bind the EGFR with varying affinity and potentially differential downstream function. They include EGF [1], transforming growth factor alpha (TGF α) [17], amphiregulin (AR) [18], heparin-binding EGF-like growth factor (HB-EGF) [19], betacellulin [20], epiregulin [21], neuregulin-2-beta (NRG2 β) [22] and the most recently discovered Epigen [23]. These peptide ligands are produced as trans-membrane precursors that are then processed by metalloproteases and released in their soluble form [24] (Fig. 1).

Ligand induced ATP binding to the EGFR lysine-721 residue is a critical step in tyrosine kinase activation and auto-phosphorylation in the intracellular region of the receptor [11, 25 - 28]. In turn, this auto-phosphorylation results in a more open

conformation allowing access to several cellular substrates to the tyrosine kinase domain of the EGFR [25, 29] and subsequent triggering of downstream signaling cascades including the RAS-RAF-MAPK-Erk1/2 pathway, the PTEN regulated phosphatidylinositol 3-kinase (PI3-K)-Akt-mTOR pathway, Src-Signal transducer and activator of transcription (STAT) family members and the Phospholipase C gamma (PLC γ) signaling pathway [30]. These signaling networks and the evidence for alterations or hyper-activity of each of these downstream molecules in providing resistance mechanisms to anti-EGFR therapy will be covered thoroughly in Part II of our series of reviews.

Due to the EGFR's many associations at the cell membrane and the diverse network of signaling, its activation is intimately associated with many cellular activities in both development and in the adult organism including proliferation, survival, differentiation, adhesion, migration and invasion and tumor metastasis. The importance of the EGFR in development is provided from the analysis of genetically altered mice. EGFR knockout mice display impaired epithelial development resulting in either embryonic or perinatal lethality or in mice suffered from abnormalities in multiple organs including the brain, skin, lung and gastrointestinal tract, depending on the genetic background [31 - 34]. Among the functions attributed to the EGFR are the proliferation and development of specific epithelial regions in the embryo, including branch point morphogenesis, maturation of early embryonic lung tissue, skin development and promoting survival of early progenitor cells in the cleft palate [35, 36]. The EGFR is also expressed throughout the brain during development primarily in the early postnatal astrocytes and purkinje cells [37, 38]. The EGFR also plays an important role in the adult organism where it is essential for the differentiation of normal mammary glands and the induction of uterine and vaginal growth [39, 40]. It is also required in the adult neurones of the cerebral cortex where it acts to promote terminal differentiation [41].

In summary these data clearly show the essential role of the EGFR during normal development and homeostasis. Not surprisingly, genetic alterations leading to EGFR over-expression or gain-of-function mutation are frequently observed in cancer [42 - 44]. These findings led to the vigorous pursuit that continues today to develop agents targeting the EGFR (and downstream substrates) in the hope that

Tumor Resistance Mechanisms to Inhibitors Targeting the Epidermal Growth Factor Receptor – Part II: Intracellular Molecules

Rodney B. Luwor*

Department of Surgery, The Royal Melbourne Hospital, The University of Melbourne, Parkville, Victoria 3050, Australia

Abstract: Tumor resistance to agents targeting the Epidermal Growth Factor Receptor (EGFR) is common, and well recognised as a major challenge to successful clinical outcome, because patients often present with tumors that contain pre-existing intrinsic resistance mechanisms to current EGFR inhibitors, which ultimately has no therapeutic benefit. Furthermore, patients who initially respond to these therapies commonly relapse, presenting with new tumors that have acquired resistance to the original therapy. Substantial translational and clinical research has been undertaken in order to understand, and more importantly overcome, the molecular initiators of both intrinsic and acquired tumor resistance. However, despite a multitude of cost and effort in gaining greater understanding of the molecular mechanisms that drive tumor resistance, very little has translated into clinical practice and management of patients. In these 2 back-to-back chapters, we will provide an overview of the progress made in targeting the EGFR and discuss the challenges presented by the numerous molecular mechanisms currently identified, leading to overall refractory outcomes to anti-EGFR therapeutics. In this chapter (Part II) we will specifically focus on the resistance mechanisms mediated by alterations in substrates downstream of the EGFR and review other intracellular mechanisms that mediate both sensitivity and resistance outcomes to anti-EGFR agents.

Keywords: Afatinib, Cancer, Cetuximab, Epidermal Growth Factor Receptor,

* **Corresponding author Rodney B. Luwor:** Department of Surgery, The Royal Melbourne Hospital, The University of Melbourne, Parkville, Victoria 3050, Australia; Tel: +613 8344 3027; Fax: +613 9347 6488; E-mail: rluwor@unimelb.edu.au

Erlotinib, Gefitinib, Lapatinib, Panitumumab, Resistance, Signaling, Therapeutics, Tumor.

1. INTRODUCTION

The HER or ErbB family consists of four members, the Epidermal Growth Factor Receptor (EGFR) (also referred to as ErbB1 or HER1) [1], HER2 (p185^{Neu} or ErbB2) [2], HER3 (ErbB3) [3] and HER4 (ErbB4) [4] and is one of the most intensely studied, and targeted, receptor tyrosine kinase families. Inactive EGFR exists mainly in a “tethered” conformation where extracellular domains II and IV associate intra-molecularly leading to an auto-inhibitory favoured state. This conformation occludes the accessibility of the dimerization arm of the receptor (in domain II) and separates two regions in domain I and III involved in ligand binding. Upon ligand binding, the ligand binding regions of domain I and III are brought closer together and the EGFR converts into an extended conformation resulting in a dis-association of the auto-inhibitory interaction of domain II and IV and exposure of the dimerization arm facilitating dimerization of the extracellular region [5 - 7].

In addition, the ligand induced extended conformation of the EGFR instigates ATP binding to a lysine residue, (Lys-721), within the EGFR kinase domain [8]. This binding is a critical event required for rapid intrinsic tyrosine kinase activation and auto-phosphorylation of specific tyrosine residues in the intracellular domain of EGFR [9 - 12]. This auto-phosphorylation in turn results in a more open conformation permitting the access of cellular substrates to the tyrosine kinase domain [8, 13]. The phosphorylated tyrosines of the EGFR serve as high affinity docking sites for Src homology 2 (SH2) and phospho-tyrosine binding (PTB) domain containing signalling proteins [14, 15]. Mutational analysis has shown that the removal of the auto-phosphorylation sites has a severe effect on substrate binding if all five tyrosine sites are removed. However, when only one site is altered, the remaining auto-phosphorylated sites appear to be able to compensate for the loss of the tyrosine site [16]. Adding to the diversity of EGFR downstream signaling is the presence of other ligands including the neuregulin family that activate the EGFR indirectly by binding HER3 and HER4 and resulting in EGFR trans-phosphorylation by EGFR-HER3 or EGFR-HER4 dimerization [17, 18].

Furthermore, the EGFR can co-operate with many other non-ErbB family members receptors leading to increased diversity of signaling pathway activation downstream [19 - 30]. It is thought that each ligand within the ErbB family illicit a subtly distinct conformation between the two dimerizing receptors in the intracellular region, resulting in differential tyrosine phosphorylation profiles and unique sets of docking substrates, ultimately leading to distinct biological outcomes [31]. Proteins that directly bind the phosphorylated tyrosines of the EGFR through their SH2 domain include PLC-(, GAP, Grb2, and Crk [15, 32 - 34] while others such as Shc interact *via* their PTB domain [32]. HER2, HER3 and HER4 also contain areas in their intracellular region for SH2 domain containing proteins to bind [34]. However, each HER receptor displays a distinct set of C-terminal auto-phosphorylation sites resulting in the recruitment of a different set of substrates. The recruitment and activation of these molecules to the receptor in turn selectively activates downstream signaling networks which include the RAS-RAF-MAPK-ERK1/2 pathway, the PTEN regulated phosphatidylinositol 3-kinase (PI3-K)-Akt-mTOR pathway, Src-Signal transducer and activator of transcription (STAT) family members and the Phospholipase C gamma (PLC γ) signaling pathway [35] (Fig. 1). In turn these signalling molecules interact with nuclear transcription factors and cytoskeletal proteins triggering gene transcription of many proteins involved in regulating a variety of cellular functions and changes in cell polarity and morphology [36, 37].

Furthermore, despite being originally recognised as a mechanism in which cells inactivate signalling by internalisation and degradation of activated receptors, EGFR signalling is sustained or initiated following receptor endocytosis and subsequent trafficking through early and late endosomes [38 - 40]. In addition, the EGFR (and the other HER family members) translocate into the nucleus where they are involved in direct gene transcription [41 - 47]. Finally, the presence of EGFR ligands, full length EGFR and the truncated variant EGFRvIII, that are all signaling competent have been discovered in secreted exosomes suggesting an inter-cellular role of EGFR signaling [48 - 50].

Not surprisingly, due to the EGFR's many associations at the cell membrane and the diverse network of signaling, and as outlined in our previous review (Part I of this series), EGFR activation is intimately associated with many cellular activities

Chemotherapeutic, Immunologic, and Molecularly Targeted Therapy for the Treatment of Advanced Melanoma

Saïd C. Azoury¹, David M. Straughan², Robert D. Bennett², Vivek Shukla^{3,*}

¹ Department of Surgery, The Johns Hopkins Hospital, Baltimore, MD, USA

² Department of Surgery, University of South Florida, Morsani College of Medicine, Tampa, FL, USA

³ Thoracic and GI Oncology Branch, National Cancer Institute, National Institutes of Health, Bethesda, MD, USA

Abstract: Over the past several decades, the incidence of melanoma has increased. Although surgery remains the primary treatment modality for localized early-stage lesions, melanoma is often diagnosed following locoregional and distant disease spread. Prognosis for advanced stage disease is dismal as one would expect; however, nowadays, the myriad of systemic therapies have allowed for improvements in disease free and overall survival. Such systemic treatment approaches include chemotherapy, immunotherapy, and molecularly targeted agents. Since the time of the approval of dacarbazine by the Food and Drug Administration for the treatment of metastatic melanoma in 1975, other agents have gained approval including interleukin-2, immune checkpoint inhibitors such as ipilimumab (anti-CTLA-4), and others. More recently, studies suggest that combination regimens of the aforementioned approaches may further improve outcomes when compared to monotherapy. Herein, the authors provide an up-to-date comprehensive review on the chemotherapeutic, immunologic, and molecularly targeted therapy approaches to the treatment of advanced melanoma.

Keywords: Advanced Stage, Anti-PD-1, Anti-CTLA-4, Anti-PD-L1, Chemotherapy, Dacarbazine, Immune Checkpoint, Immunotherapy, Interleukin-2,

* **Corresponding author Vivek Shukla:** 10 Center Drive, Room 3W5848, Thoracic and GI Oncology Branch, Clinical Center, National Cancer Center, National Institutes of Health, Bethesda, MD-20892, USA; Tel: 301-594-8961; E-mails: shuklav@mail.nih.gov, vivek.shukla@nih.gov

Melanoma, Metastatic, Systemic, Targeted Therapy, Vaccine.

INTRODUCTION

Over the past several decades, the incidence of melanoma has markedly increased [1 - 3]. Worldwide, approximately 50,000 deaths can be attributed to melanoma each year [4]. Melanomas originate from melanocytes, which reside in the basal layer of the epidermis and produce melanin. Several mechanisms are thought to underlie the malignant transformation of normal melanocytes in melanoma development. Perhaps one of the most well studied risk factors for melanoma development is ultraviolet radiation [5]. Under normal circumstances, melanin helps protect the skin from ultraviolet light. Overtime, excessive sun exposure, results in excessive DNA damage to proliferating melanocytes, thereby overwhelming the normal DNA repair mechanisms. Occasionally in the process, a cell will undergo malignant transformation [5]. Furthermore, several genes, such as *BRAF*, *PTEN*, *c-Kit*, *p53*, *CDKN2A/p16* are implicated in the development by mutation, deletion, or amplification. Mutations in these genes may be induced by UV radiation or are inherited and ultimately result in the dysregulation of the normal cell cycle checkpoints. Understanding the complex interplay of genetic and environmental factors in melanoma pathogenesis is an ongoing area of investigation.

Surgery is the primary treatment modality for localized early stage lesions, with estimated 5 and 10-year survival rates approximating 97% and 93% for patients with Stage 1a, T1aN0M0 melanomas ($\leq 1\text{mm}$, without ulceration and mitosis $\leq 1/\text{mm}^2$) [6]. Metastatic melanoma carries an overall median survival of 4-12 months, depending on the site of distant disease [7]. The current melanoma TNM staging system was recently updated using data from an expanded American Joint Committee on Cancer (AJCC) Melanoma Staging Database [4]. For metastatic melanoma (stage IV), elevated serum levels of LDH and site(s) of metastases define the M1 stage into three categories: M1a - only distant skin, subcutaneous or nodal metastases and normal LDH, M1b - lung metastases is present with a normal LDH, or M1c - metastases to any other visceral site and elevated LDH [4]. As one would expect, prognosis worsens as disease progresses from M1a to M1c disease.

While the prognosis after surgical resection in patients with early stage disease is favorable, the median overall survival for patients with distant metastases (stage IV) treated with chemotherapy is less than a year, and 5-year survival approximates 10% [3]. When considering metastatic disease, surgery may improve outcomes, and complete resection has been shown to improve survival when compared to incomplete resection [8, 9]. Melanoma is generally considered to be a relatively radio-resistant tumor; however, radiation therapy (*e.g.*, whole brain irradiation or stereotactic radiosurgery) has been used in adjuvant and palliative settings, such as in cases of metastasis to the brain [10 - 12].

Table 1. Select agents approved by the US Food and Drug Administration (FDA) for the treatment of advanced melanoma.

Class	Agent	Year FDA Approved	Indication
Chemotherapy	Dacarbazine	1975	Stage IV melanoma
Immunotherapy	High-Dose Interferon alfa-2b (IFN- α)	1996	Adjuvant treatment of intermediate and high-risk melanoma (Stage IIB/C, Stage III)
	High-Dose Interleukin-2 (IL-2)	1998	Stage IV melanoma
	Ipilimumab	2011	Unresectable Stage III or Stage IV melanoma
	Nivolumab	2014	Unresectable Stage III or Stage IV melanoma
	Pembrolizumab	2014	Unresectable Stage III or Stage IV melanoma
Molecularly Targeted Therapy	Vemurafenib	2011	Patients with BRAF V600E mutation with unresectable Stage III or Stage IV melanoma
	Trametinib	2013	Patients with BRAF V600E/V600K mutation who have unresectable Stage III or Stage IV melanoma
	Dabrafenib	2013	Patients with BRAF V600E mutation with unresectable Stage III or Stage IV melanoma
	Trametinib/ Dabrafenib Combination	2014	Patients with BRAF V600E/V600K mutation who have unresectable Stage III or Stage IV melanoma

Systemic therapy remains the primary treatment modality for stage IV disease [13]. Prior to 2011, high dose interleukin-2 (IL-2) and an alkylating agent,

Targeting the Warburg Effect for Cancer Therapy: A Long and Winding Road

Patrícia L. Abreu*, Ana M. Urbano*[#]

Unidade de Química-Física Molecular and Departamento de Ciências da Vida, Faculdade de Ciências e Tecnologia, Universidade de Coimbra, Coimbra, Portugal

Abstract: In the 1920s, Otto Warburg, one of the leading biochemists of the 20th century, uncovered a striking phenotype of cancer cells: their increased dependence on lactic acid fermentation for energy production compared to that of the normal cells from which they derived. Warburg viewed this metabolic particularity of cancer cells, which came to be known as the Warburg effect, as a driving force in carcinogenesis. This perception suggested a novel path for cancer therapy, a strategy that Warburg himself proposed and defended with passion to his death. However, for many decades, both his metabolic theory of cancer and suggested therapeutic approach were essentially ignored by cancer researchers, who were mostly focused on the genetic basis of the disease and on the intricacies of the pathways known to promote cellular proliferation, differentiation and death. Still, thanks to the combined efforts of those who chose to pursue Warburg's line of research, experimental evidence supporting and extending Warburg's findings on the metabolism of cancer cells accumulated. In the 1980s, ¹⁸F-deoxyglucose positron emission tomography (¹⁸FDG-PET) was implemented in the clinic. This metabolic imaging technique, which is based on the avidity of cancer cells for glucose, represents, to this day, the only successful exploitation of the Warburg effect for medical purposes. The wide success of ¹⁸FDG-PET in the diagnosis and staging of tumors is among the factors most responsible for renewing interest in the central carbon metabolism of cancer cells. This renewed interest was further boosted by the discovery of multiple links between central carbon metabolism and cellular proliferation, differentiation and death and culminated

* **Corresponding authors Ana M. Urbano and Patrícia L. Abreu:** Department of Life Sciences and Molecular Physical Chemistry Research Unit, University of Coimbra, Coimbra, Portugal; Tel/Fax: +351-239-826541; E-mails: amurbano@ci.uc.pt & patricia.lonaabreu@gmail.com

[#] Centro de Investigação em Meio Ambiente, Genética e Oncobiologia (CIMAGO), Faculdade de Medicina, Universidade de Coimbra, Coimbra, Portugal.

in the recent classification, by Weinberg and Hanahan, of tumor metabolism as an emerging cancer hallmark. Tremendous research effort is now being devoted into a more detailed and comprehensive elucidation of the metabolic rewiring that accompanies neoplastic transformation and, unsurprisingly, targeting the metabolic peculiarities of tumors has become a hot topic in drug discovery. This chapter summarizes past and current efforts at targeting the Warburg effect for selective cancer therapies.

Keywords: Aerobic glycolysis, Central carbon metabolism, Clinical trials, Diabetes, Emerging cancer hallmark, ^{18}F -Deoxyglucose positron emission tomography (^{18}FDG -PET), Hypoxia, Ketogenic diet, Metabolic cancer therapies, Metformin, Pasteur effect, Targeted cancer therapies, Warburg effect.

1. THERAPEUTICAL APPROACHES TO CANCER: FROM CHEMOTHERAPY TO TARGETED THERAPIES

The oldest written description of cancer known to exist can be found in the Edwin Smith Papyrus, which is based on what was known in surgery and medicine up to 3000 BC [1, 2]. But humans must have, in all likelihood, been fighting against cancer throughout their existence. Although this papyrus describes the treatment of tumors with cauterization, it also acknowledges the absence of a cure for the disease. By 400 BC, Hippocrates, the “Father of Medicine”, advised against the treatment of deep-seated tumors, as this would shorten the lives of patients [2]. Fortunately, this perception of cancer as an incurable disease did not prevent significant advances in cancer therapy, mostly during the last century. Some milestones in cancer therapy will be briefly discussed in this section.

The first successful inductions of tumor regression *via* systemic administration of chemical substances can be traced back to the 1940s. At that time, our understanding of human cancer biology was very limited and the discovery of the anticancer activity of the first cancer drugs stemmed from chance observations. Namely, from the post-mortem observation of severe myelosuppression and lymphoid hypoplasia in First World War soldiers dying of mustard gas exposure, which suggested the use of nitrogen mustards for the treatment of lymphomas [3, 4], and the observation of increased proliferation of acute lymphoblastic leukemia (ALL) cells upon administration of folic acid to children with ALL [5],

which suggested the use of two folate analogues (aminopterin and amethopterin (methotrexate; brand names AbitrexateTM and BrimexateTM)¹) to treat this neoplasia [6]. Although brief, due to development of tumor resistance to the drugs, these remissions stimulated further research on cancer therapy [7].

By this time, attempts were also being made at the rational design of compounds capable of interrupting cell proliferation. These early attempts led to the development of several purine analogs, designed to interfere with the natural production of DNA. The two most promising analogs, 6-mercaptopurine (PurinetholTM, Alti-MercaptopurineTM) and thioguanine (TabloidTM, LanvisTM), were introduced in the clinic in the 1950s, establishing a novel class of cancer drugs [8, 9]. Notwithstanding these attempts at rational drug design, serendipity continued to play a role in the discovery of novel classes of cancer drugs. That was the case with vinca alkaloids, whose anticancer potential emerged in a screen for antidiabetic activity [10]. Other discoveries were the result of systematic screenings, most notably that conducted on thousands of natural products by the National Cancer Institute, which led to the discovery of the anticancer activity of taxanes and camptothecins [11].

Once the mechanisms of action of these cancer drugs became known, new compounds with similar actions, but with refined structures, could be synthesized. It was hoped that these novel structures would improve their pharmacological properties, namely in terms of stability and efficacy. However, gains achieved using this approach were rather modest [7]. Those achieved through further systematic screenings were, likewise, modest, as most of the new cancer drugs thus discovered belonged to the classes already in clinical use [12].

It is worth noting that, in spite of exhibiting distinct mechanisms of action, all cancer drug classes used in the clinic up to the mid 1970s acted by directly interfering with cellular proliferation: nitrogen mustards are non-specific DNA alkylating agents [13]; the antimetabolite methotrexate inhibits the enzyme dihydrofolate reductase (DHFR), thereby compromising the synthesis of thymidine and purines and, ultimately, DNA synthesis [14]; vinca alkaloids and taxanes are both antimitotic agents [15]; camptothecin is an inhibitor of topoisomerase I [16], an enzyme essential for DNA unwinding during replication

Immunotherapy Strategies in Follicular Lymphoma: Antibodies, Vaccines and Cells

Nicolás Martínez-Calle¹, Ascensión López Díaz de Cerio¹, Susana Inogés¹, Esther Pena³, Ricardo García-Muñoz², Carlos Panizo^{1,*}

¹ Hematology Department, Clínica Universidad de Navarra, Pamplona, Spain

² Hematology Department, Hospital San Pedro, Logroño, Spain

³ Hematology Department, Complejo Hospitalario de Navarra, Pamplona, Spain

Abstract: Follicular lymphoma (FL) is the most frequent indolent non-Hodgkin lymphoma. Therapeutic strategies vary from withholding treatment to aggressive chemoimmunotherapy regimens, and stem cell transplantation, depending on the stage and risk stratification at diagnosis. A prominent role of the microenvironment in FL-cell survival and lymphomagenesis has been brought to light and consequently the manipulation of the FL-cell niche is progressively becoming an important therapeutic tool in FL. Chemotherapy agents are no longer under the spotlight, leaving the main role to immunotherapeutic strategies and targeted therapy that aim towards disease control with minimal side-effects and sequelae. Immunotherapy with monoclonal antibodies, radioimmunotherapy and vaccines, has resulted in increased response rates and survival in FL patients.

Adoptive immunotherapy is an emerging strategy for FL treatment, aiming to exploit the immune system's natural tendency to attack tumoral cells. AntiCD20 monoclonal antibodies have become the backbone of first line and relapse treatments combined with chemotherapy regimens. Anti-idiotypic vaccines are the best developed active immunotherapy strategy, with proven efficacy in patients with FL on first relapse. The other vaccine types (Dendritic cells, proteoliposomal or DNA) are still in preclinical development. Adoptive cell transfer (NK cells, LAK and effector T-lymphocytes), chimeric-antigen receptor (CAR) engineered T-cells and Bi-specific T-cell engaging

* **Corresponding author Carlos Panizo:** Hematology Department, Clínica Universidad de Navarra, Av. Pío, XII, 36 31008 Pamplona, Navarra, Spain; Tel: +34 948 396 397; Fax: +34 948 396 676; Email: cpanizo@unav.es

antibodies (BiTE) for passive immunotherapy remain also experimental approaches, although promising pre-clinical results have recently become available.

The following chapter will summarize FL biology and conventional treatment with immunochemotherapy, with a final section focusing specifically on novel immunotherapy strategies (active and passive) for the treatment of FL.

Keywords: Adoptive cell-therapy, Anti-idiotypic vaccines, Dendritic-cell vaccines, Follicular lymphoma, Immunotherapy, LAK cells, Monoclonal antibodies, NK cells, Radioimmunotherapy, Vaccines.

1. INTRODUCTION

Follicular lymphoma (FL) is the most frequent indolent non-Hodgkin's lymphoma (NHL), it is known to arise from the follicular B lymphocytes and typically features an indolent clinical course consisting of relapses followed by prolonged remissions. FL was originally named Brill-Symmers disease, it was first described in 1925 as a benign adenopathic disorder, typical of the elder [1]. FL is the second most common NHL in United States and Eastern Europe, representing 20-40% of all NHL and 70% of indolent lymphomas, with a yearly incidence of 3/100.000 [2, 3]. Definitive cure of FL seems to occur rarely [4], although patient survival continues to improve with a current average above 10 years [5 - 8].

Improvements in the outcome of FL have been driven by the appearance of novel biologic agents [9] together with a better risk assessment at diagnosis, which combines the traditional FLIPI (Follicular Lymphoma International Prognostic Index) [10] with genetic and molecular biomarkers [11]. These tools have helped clinicians to individualize treatment intensity, reduce unnecessary treatment-related toxicity and ultimately achieve durable complete remissions [12, 13]. Immunotherapy (*e.g.* anti CD20 antibodies, radioimmunotherapy or idiotypic vaccination) have probably been the greatest advance in FL treatment over the past 50 years, contributing to the extension of disease free intervals and challenging some of the oldest paradigms about FL treatment, such as incurability or the use of front-line aggressive treatment to extend survival [14].

2. BIOLOGY OF FOLLICULAR LYMPHOMA

2.1. Histopathology of FL

FL is a neoplasm composed of germinal center B cells (also known as follicle center cells), typically both centrocytes and centroblasts, maintaining at least partially the follicular histologic pattern. FL is characterized by the t(14;18)(q32;q21) translocation resulting in overexpression of anti-apoptotic BCL2 protein [15 - 17]. The World Health Organization (WHO) classification of malignant lymphomas defines three histologic grades of FL (1, 2 and 3), based on the number of centroblasts per high power field, using the cell counting method developed by Mann and Bernard [18]. More than 15 centroblasts per high power field defines grade 3 which is subsequently classified into grades 3a and 3b (3a representing an admixture of centrocytes and centroblasts and 3b predominant sheets of centroblasts). Grade 3 FL is heterogeneous, with increasing aggressiveness and less survival correlating with increasing centroblast counts [19]. In fact, clinical course of grade 3b FL overlaps with that of aggressive NHL and therefore should be treated as such [15]. FL tumor cells express surface immunoglobulin (SIg+: IgM, IgD, IgG or rarely IgA) and also express typical B cell associated antigens (CD19, CD20, CD22, CD79a). Other phenotypic features include BCL2+, BCL6+, CD10+, CD5-, CD43+/- [20, 21]. Some cases, especially grade 3b, may lack CD10 but retain BCL6 expression. CD43 is a common marker of grade 3 FL [22, 23].

2.2. Follicular Lymphoma-cell Origin

The t(14;18)(q32;q21) translocation involves BCL2 and IGH genes, the breakpoint is located at the 5' end of J heavy-chain (JH) gene, suggesting that the event occurs at the DH to JH rearrangement stage; such event occurs in bone marrow lymphoid progenitors or B cell precursors (pro-B and pre-B cells) [24 - 28]. Furthermore, FL-cells occasionally display class-switch recombination (CSR), suggesting altogether an origin between lymphoid progenitors in bone marrow and late germinal center follicular cells [29, 30].

After the immortalization event, FL-cells are thought to continue their normal differentiation path through the germinal center in spite of the BCL2-IGH

Epigenetics and Cancer Cell Metabolism: Cross-talk and Therapeutic Opportunities

Chi Chun Wong*, Jun Yu*

Department of Medicine and Therapeutics, State Key Laboratory of Digestive Disease, Li Ka Shing Institute of Health Sciences, Shenzhen Research Institute, The Chinese University of Hong Kong, Hong Kong

Abstract: Epigenetics is increasingly recognized to play an important role in tumorigenesis. The epigenome encompasses a multitude of elements that regulate gene expression, including DNA methylation, histone modification, microRNA, and more recently, non-coding RNA. Aberrant regulation of the epigenome has been implicated in altered gene expression and function, which contribute to cancer development and progression *via* the promotion of cellular transformation, metastatic spread, and drug resistance. Emerging evidence indicates that the activities of key epigenetic regulators including DNA methyltransferases and histone modification enzymes are sensitive to cellular metabolism. The efficiency of these metabolic enzymes depends on the availability of substrates and/or co-factors that can be profoundly altered in cancer. Mutations in metabolic enzymes in cancer also generate oncometabolites that can lead to the dysfunction of DNA and histone demethylases. Conversely, through mediating aberrant expression of genes that are involved in cellular metabolism, epigenetic mechanisms could contribute to metabolic rewiring in cancer to confer a growth advantage to cancer cells. Understanding this cross-talk between epigenetics and cancer cell metabolism may unravel novel therapeutic opportunities. In this chapter, we will review recent discoveries linking epigenetics and cancer cell metabolism, their implications in oncogenesis, and highlight potential approaches to target these cancer-specific abnormalities therapeutically.

* **Corresponding authors Chi Chun Wong and Jun Yu:** Institute of Digestive Disease and Department of Medicine and Therapeutics, Prince of Wales Hospital, The Chinese University of Hong Kong. Shatin, NT, Hong Kong; Tel: (852) 3763 6099; E-mails: chichun.wong@cuhk.edu.hk; junyu@cuhk.edu.hk

Atta-ur-Rahman (Ed.)

All rights reserved-© 2016 Bentham Science Publishers

Keywords: Acetyl-coenzyme A, Cancer, Cancer metabolism, DNA methylation, Epigenetics, Fumarate hydratase, Gene expression, Glutaminolysis, Glycolysis, Histone acetylation, Histone demethylase, Histone methylation, Isocitrate dehydrogenase, MicroRNA, Mitochondrial succinate dehydrogenase, Non-coding RNA, S-adenosylmethionine, TET methyl-cytosine dioxygenase, Tricarboxylic acid cycle, Warburg hypothesis.

INTRODUCTION

Epigenetics is defined as heritable changes in gene expression that are not resulted from change(s) in the underlying DNA sequence. Epigenetic regulation of gene expression can be highly dynamic and of a transient nature, or can be relatively stable and be passed to offspring through the germline. Given its role in the regulation of gene expression, it is increasingly recognized that epigenetics play an important role in cell growth, differentiation and development. The human ‘epigenome’ encompasses three major elements that interact with each other to co-operatively to either activate or silence gene expression, which involves direct chemical modification of DNA by methylation, alteration of DNA accessibility by histone modifications, and the selective silencing of mRNA levels by noncoding RNA.

The tight regulation of the epigenome is essential in normal cellular processes. Consequently, disruption of the epigenetic machinery can cause the inappropriate activation or silencing of genes, leading to the development of numerous diseases. The notion that epigenetic disruptions may be associated with cancer development was first proposed in the 1980’s [1, 2]. Extensive research over the past two decades has clearly demonstrated that epigenetic dysregulation contributes to tumorigenesis by silencing of tumor suppressor genes [3 - 5]. With the advent of next-generation sequencing in conjunction with chromatin immunoprecipitation (ChIP-Seq) [6] and microarray technologies (*e.g.* Illumina 27K and 450K) [7], we are just beginning to appreciate the impact of epigenetic dysregulation in the development of cancers on a genome-wide scale [8].

Comprehensive molecular characterization of the cancer genome, transcriptome, epigenome and proteome by the Cancer Genome Atlas (TCGA) has further

highlighted the role of aberrant epigenetics across different types of cancer, such as promoter DNA hypermethylation in a subset of colorectal cancers [9] and extreme hypermethylation in Epstein-Barr virus (EBV)-associated gastric cancer [10]. Moreover, whole genome sequencing has unraveled a number of epigenetic regulators, such as chromatin modification enzymes, that are recurrently mutated in various cancers, indicating that they may be driver mutations in carcinogenesis [11].

Recent studies have revealed an intricate relationship between epigenetics and cancer metabolism [12 - 14]. Cancer cells exhibit metabolic alterations to support an increased biosynthesis [15] and adaptations to allow their proliferation under an adverse microenvironment [16]. For example, cancer cells consume more glucose than normal cells through aerobic glycolysis, an inefficient pathway that generates much less ATP than oxidative phosphorylation, a phenomenon also known as the ‘Warburg’s effect’ [17]. Such a metabolic re-programming is crucial for cancer cell survival, and is activated by oncogenic signaling cascades such as PI3K-AKT-mTOR [18] and transcription factors such as the hypoxia-inducible factor (HIF) and MYC [19], and the inactivation of tumor suppressor signaling, *e.g.* LKB-AMPK [20].

Metabolic sensing by the epigenetic machinery represents a built-in mechanism to regulate cellular activities in response to environmental clues, and it is frequently deregulated in cancers [12, 16]. There exists a four-way cross-talk between epigenetics and metabolism in cancer: epigenetic dysfunction that 1) directly affects expression of metabolic enzymes; and 2) indirectly alters signaling transduction cascades involved in the control of cell metabolism; and metabolic alterations that 3) influence the availability of substrates and cofactors necessary for the proper functioning of epigenetic modification enzymes; and 4) result in the production of oncometabolites that act as agonists or antagonists for epigenetic modification enzymes. In this chapter, we will provide a brief background on the epigenetic and metabolic dysregulation in human cancers, followed by a detailed account of the cross-talk between epigenetics and cancer metabolism, its underlying cause, clinical significance and potential therapeutic implications.

Apoptosis Targeting Therapeutics in Clinical Trials

N.S. Hari Narayana Moorthy^{1,2,*}, C. Karthikeyan², Piyush Trivedi²

¹ Departamento de Química e Bioquímica, Faculdade de Ciências, Universidade do Porto, s/n, Rua do Campo Alegre, 4169-007 Porto, Portugal

² School of Pharmaceutical Sciences, Rajiv Gandhi Proudyogiki Vishwavidyalaya, Airport Bypass Road, Gandhi Nagar, Bhopal (MP)-462033, India

Abstract: Apoptosis (called as programmed cell death) is vital for maintaining homeostatic balance between cell survival/cell deaths in metazoan cells. Apoptosis is regulated through extrinsic (or receptor mediated) and intrinsic (or mitochondria mediated) pathways. The pro-apoptotic proteins (e.g. Bax, Bak, Bad, Bcl-Xs, Bid, Bik, Bim and Hrk) and the anti-apoptotic proteins (e.g. Bcl-2, Bcl-XL, Bcl-W, Bfl-1 and Mcl-1) are crucial to control the apoptotic pathways. Dysfunctions of apoptosis pathways are implicated in cancer as defects in these pathways not only promote tumorigenesis but also confer resistance to cancer cells to most conventional chemotherapies as well as radiotherapy. The apoptosis occurs by imbalanced pro-apoptotic and anti-apoptotic protein levels, impaired or reduced death receptor signalling and caspase function. Hence, targeting apoptosis pathways is considered as an attractive strategy for therapeutic intervention in cancer. The past decade recorded tremendous advances in this area especially small molecular intervention of apoptosis pathways for cancer treatment which resulted in several compounds under clinical development. This chapter reviews the current progressions in the development of bioactive molecules targeting apoptotic pathways with special emphasis on small molecular anticancer drugs under clinical trials. Some excellent examples are; nutlins, MI-888, MI-219 and SM-164 which target MDM2, ABT-263, AT-406 and GX15-070MS which target Bcl-2 family of proteins, birinapant, GDC-0917, HGS-1029 and LCL-161 which target IAPs (inhibitors of apoptotic proteins). The content of this chapter will be enlightening the readers in academic and research to update their knowledge on the anticancer drugs especially target proteins responsible for apoptosis.

* **Corresponding author N.S. Hari Narayana Moorthy:** Departamento de Química e Bioquímica, Faculdade de Ciências, Universidade do Porto, s/n, Rua do Campo Alegre, 4169-007 Porto, Portugal; Tel: +917552678883; Fax: +917552742002; E-mails: hari.moorthy@fc.up.pt, hari.nmoorthy@gmail.com

Atta-ur-Rahman (Ed.)

All rights reserved-© 2016 Bentham Science Publishers

Keywords: Apoptosis, BCL family proteins, Cancer, Caspase, Clinical trials, IAP, MDM2, Nutlins, p53, Pro-apoptotic protein.

CANCER

Cancer is a life threatening, multifaceted disease that involves disruption of the normal balance of cellular life and death through dysregulation of cellular homeostasis and the prevailing mechanisms responsible for cell growth and replication. Mechanisms responsible for cancer development include dysregulated response to growth signals, angiogenesis, uncontrolled replication, tissue invasion and metastasis, and evasion of apoptosis. Statistically, cancers accounted for 8.2 million deaths and 14 million new cases in 2012 and the number of cancer related deaths is expected to rise to 22 million within the next two decades [1 - 3].

Anticancer drug discovery is a challenging task owing to high attrition rate mainly attributed to lack of efficacy, non-selectivity, toxicity and incompatible pharmacokinetic profiles of anticancer agents which are under clinical development. Hence, discovery of “safe and effective” anticancer drugs remains a priority area for researchers working on cancer worldwide. The effectiveness of drugs used for the cancer therapy is dependent on the type of targets which it modulates. Biochemical pathways necessary for growth and survival of cancer cells are attractive targets for anticancer drug discovery. The former comprises mainly of signal transduction pathways regulating growth and proliferation of cancer cells while the latter comprises of pathways which enhances survival of cancer cells by imparting ability to repair and evade cell death (apoptosis). Targeting apoptosis pathways for anticancer drug discovery is relatively a new avenue compared to the much established therapeutic strategies targeting pathways regulating growth and proliferation of cancer cells. Nevertheless, the increased understanding of molecular mechanisms that regulate apoptosis together with convincing proof-of-principle evidence obtained in several animal models confirming the validity of apoptosis targeted drug discovery for cancer led to the development of several apoptosis-based therapeutics for cancer therapy. This chapter reviews the advancements in the discovery and development of small-molecules attacking apoptotic pathways with special emphasis on the bioactive molecules under clinical trials.

APOPTOSIS

Apoptosis (programmed cell death) is essential for maintaining homeostatic balance between cell survival/cell deaths in metazoan cells. The role of apoptosis in the physiological and the pathological conditions remains to be an intensely investigated area in biological research [4]. Apoptosis is triggered by imbalance in the pro-apoptotic and anti-apoptotic protein levels resulting from impaired or reduced death receptor signalling and caspase functions. Apoptosis is accompanied by a series of biochemical changes including caspases activation, DNA and protein breakdown, membrane changes and recognition by phagocytic cells [5, 6]. Dysfunction of apoptosis pathways are implicated in cancer as defects in these pathways not only promote tumorigenesis but also confer resistance to cancer cells to most conventional chemotherapies as well as radiotherapy. Hence, targeting the apoptosis pathways is considered as an attractive strategy for therapeutic intervention in cancer [7].

MECHANISMS OF APOPTOSIS

Understanding the mechanism of apoptosis formation is vital to comprehending the pathogenic circumstances developed from disordered apoptosis (Fig. 1) [5]. Apoptosis is triggered either through mitochondria (intrinsic) or death receptor mediated pathways (extrinsic). A myriad of stress signals caused by therapeutics (chemo and radiotherapies) activate the intrinsic pathways of the apoptosis. Subsequently, the signal is relayed to the mitochondria upon the stress, leading to the mitochondrial outer membranes permeabilization (MOMP). This allows the apoptotic proteins including cytochrome c and second mitochondrial-derived activator of caspases (SMAC) to be released into the cytosol from mitochondria. The cytochrome c causes the formation of a multiprotein complex (apoptosome)-cytochrome c, apoptotic protease activating factor 1 (APAF1) and procaspase-9, initially. This causes activation of the caspase-9 activity and downstream caspase cascade from procaspase-9. Further, the release of SMAC stimulates caspase activation by neutralizing IAPs, which regulates apoptosis through inhibition of caspases [5, 6].

SUBJECT INDEX

A

- Acetylation 390, 402, 403, 409, 410, 412
- Acetyl CoA 278, 279, 281, 303, 306, 410, 411, 415
- Acetyl-coenzyme A 387
- Acetyl group 391, 411
- Activation of AMPK 396, 401, 408
- Activity 13, 272, 273, 304, 309
 anticancer 272, 273, 304
 significant 13, 309
- Acute myeloid leukemia (AML) 286, 293, 301, 349, 351, 390, 416, 419, 465, 475, 476, 477
- Adenocarcinomas 41, 292, 293, 295
- Adoptive cell-therapy 326
- Advanced lung cancer 14
- Advanced melanoma 221, 227, 231, 234, 236, 237, 241, 242, 243, 244, 245
- Advanced NSCLC 14, 15, 31, 33, 63, 65, 302
- Advanced solid tumors 295, 296, 299, 300, 304, 463, 470, 471, 476
- Aerobic glycolysis 272, 277, 282, 283, 284, 286, 310, 311, 388, 394, 406
- Aerobic respiration 277, 278, 280, 299, 305, 311
- Afatinib 3, 9, 18, 45, 51, 64, 79, 142, 145, 185
- Agios pharmaceuticals 294, 295, 296
- Alpha-ketoglutarate 286, 313, 395, 416, 417, 418, 420, 421, 422
- Alkylating agents 219, 220, 222, 336
- American joint committee on cancer (AJCC) 218
- Amino acids 4, 40, 54, 391, 394, 413, 415
- Amphiregulin 4, 20, 81, 188
- AMPK activity in cancer cells 396
- Anderson cancer center 289, 290, 291, 292, 293, 297
- Anthracycline 13, 290, 335
- Anti-apoptotic proteins 454, 458, 460, 472
- Anticancer 273, 288, 299, 301, 305, 308, 309
- Anticancer drug discovery 455, 479
- Anticancer drugs 303, 454, 466, 479
- Anti-EGFR agents 26, 29, 34, 48, 53, 62, 66, 68, 76, 77, 142, 147, 164, 181, 187
- Anti-EGFR inhibitors 9, 10, 56, 59, 69, 74, 75, 158, 159, 175, 176, 181, 183, 185, 186
- Anti-EGFR therapeutics 3, 9, 80, 142, 164, 186
- Anti-EGFR therapy 19, 20, 43, 54, 57, 61, 63, 64, 68, 73, 146, 150, 159, 166, 175
 resistance mechanisms to 5, 164
- Antigen-presenting cells (APCs) 234, 236
- Antigens 230, 234, 350, 351, 353, 354, 356, 358, 359
 cancer-testis 230, 232
- Anti-idiotype vaccines 325, 326
- Antitumor activity 348, 461, 471, 472, 476
- Apoptosis 10, 285, 454, 455, 456, 457, 458, 464, 465, 466, 467, 468, 474, 475, 477, 478, 479
 formation 456, 457

Autologous stem cell transplantation (ASCT) 333, 338, 340, 342, 343, 344
AXL expression 77, 78, 79

B

Bacilli calmette-guerin (BCG) 233, 235
BCRP expression 148
Benign prostatic hyperplasia (BPH) 288, 289, 299, 300
Bevacizumab 75, 76, 77, 161, 225, 275
Biopsies 49, 50, 52, 175, 178, 179, 186
 patient-matched 174
BIR domains 467, 468, 469
Block ligand binding 7, 9
Bone marrow (BM) 327, 330, 331
Bortezomib 342, 343, 474
B-RAF and PIK3CA mutations 57, 183
BRAF mutations 161, 225, 242, 247, 249, 250
B-RAF mutations 160, 161, 162, 163, 164, 170
 mCRC patients harbouring 162, 163
 patients harbouring 161
B-RAF mutations in mCRC patients 161
BRAF V600E mutation 161, 219, 249, 250
BRAF V600E/V600K mutation 219
Brain metastases 13, 221, 223, 224, 232
Breakpoint cluster region (BCR) 276, 328, 329
Breast cancer 29, 59, 60, 61, 79, 148, 172, 181, 252, 274, 275, 289, 294, 297, 301, 303, 308, 309, 390, 400, 402, 472
 metastatic 29, 274, 301, 303
 cell lines 60, 61, 79, 148, 172, 181, 472
Breast cancer patients 13, 24

BTC and EPGN in mCRC patients 28
BTC expression 28

C

Cancer cell lines 148, 401, 403, 404, 464, 469, 471, 479
Cancer cells 23, 24, 53, 60, 74, 147, 184, 271, 274, 275, 277, 282, 283, 284, 288, 305, 311, 388, 394, 395, 396, 397, 407, 411, 416, 419, 424, 455, 464
 acquired resistant PC3 prostate 74
 avidity of 271, 284
 breast 23, 60, 424
 cetuximab-refractory DIFI colon 53
 colon 60, 147, 184, 407
 gefitinib-resistant breast 24
 lung 184, 424, 464
 phenotype of 271, 282
 proliferation of 419, 455
 resistant 474
 resistant H226 NSCLC 79
 survival of 388, 455
 well-aerated 277, 282
Cancer development 307, 386, 387, 413, 428, 455
Cancer drugs 272, 273, 312
Cancer hallmark, emerging 272
Cancer metabolism 307, 387, 388, 393, 397, 404, 423, 424, 427
Cancers 59, 66, 241, 243, 244, 272, 293, 301, 386, 388, 389, 391, 395, 396, 400, 402, 404, 405, 406, 410, 414, 455
 colorectal 59, 66, 241, 243, 244, 293, 301, 388
Cancer-testis antigens (CTAs) 230, 232, 233
Cancer treatment 81, 187, 239, 275, 288, 307, 427, 454, 459
Cancer vaccines 233, 275

- Capecitabine 13, 24, 76, 172, 179
 Cappuzzo 30, 35, 36, 58, 63, 71
 Carboplatin 12, 33, 35, 45, 158, 222, 295
 Carboplatin/ paclitaxel (CP) 224, 225
 Carboplatin/paclitaxel chemotherapy in patients 297
 Carcinogenesis 251, 271, 283, 284, 285, 297, 312, 388, 389, 399, 414, 416, 427
 Caspase-8 458, 466, 469
 Castration-resistant prostate cancer (CRPC) 179, 292, 304
 Cell lines 8, 13, 18, 20, 24, 25, 27, 28, 43, 52, 58, 60, 61, 66, 67, 69, 70, 73, 147, 148, 151, 159, 165, 171, 174, 180, 184, 185, 420, 426, 464, 477
 acquired gefitinib NSCLC 61, 66
 cetuximab-sensitive parental 58, 60
 gefitinib-acquired resistant breast cancer 69
 gefitinib-resistant 147
 gefitinib-resistant breast cancer 148
 gefitinib-resistant MDA-MB-468 148
 gefitinib-sensitive 70, 148
 gefitinib-sensitive parental 67, 69, 73, 147
 lung cancer 184, 464
 prostate cancer 28, 69
 resistant 52, 58, 70, 151, 159, 171, 174, 477
 trastuzumab-resistant breast cancer 13
 Cell membrane 4, 5, 144, 147, 148
 Cells 22, 23, 60, 70, 75, 147, 148, 149, 153, 185, 280, 328, 329, 330, 355, 454, 456
 acquired cetuximab-refractory DIFI 53
 cetuximab-resistant 149
 cetuximab-resistant DiFi 149
 cetuximab-sensitive 149
 erlotinib-resistant 147
 gefitinib-resistant 147
 gefitinib-resistant A431 70, 148
 gefitinib-sensitive 60
 gefitinib-sensitive prostate PC3 75
 metazoan 454, 456
 muscle 280, 355
 non-tumoral 328, 330
 parental A431 22, 23
 refractory 70, 149, 185
 stromal 328, 329
 Cell types 278, 279, 307, 328, 330, 331, 348, 407
 Cellular proliferation 247, 271, 273, 479
 Cetuximab 10, 11, 12, 22, 24, 27, 33, 34, 35, 42, 52, 53, 54, 57, 65, 67, 71, 79, 149, 154, 155, 158, 159, 161, 163, 164, 165, 166, 168, 169, 174, 183
 addition of 155, 164
 combined 24, 42
 comparing 10, 12
 evaluating 11, 54, 165
 following 71, 158, 168
 mechanism of resistance to 166, 183
 post 52, 53, 67, 174
 single-agent 12, 33
 Cetuximab 7, 9, 10, 11, 12, 21, 22, 24, 25, 26, 27, 28, 47, 50, 56, 57, 60, 62, 63, 66, 71, 74, 75, 76, 79, 149, 151, 152, 153, 154, 155, 156, 159, 162, 165, 168, 169, 176, 178,
 -acquired resistance 149
 and irinotecan 11, 21, 27, 62, 76, 156
 and panitumumab 9, 22, 50, 155, 159
 cetuximab 152, 153, 162, 163, 168, 169, 176
 efficacy 22, 25, 60, 74, 149, 178
 /erbitux 7, 10

Subject Index

cetuximabf 153, 162, 169, 176
cetuximabg 162, 169
infusion 27
inhibition 47, 56, 66
monotherapy 10, 11, 12, 21, 22, 26,
28, 57, 63, 71, 75, 154
monotherapy group 11
resistance 60, 165
 displayed increased 24
 response 11, 21, 56, 79, 151
Cetuximab treatment 25, 27, 32, 33, 34,
55, 59, 60, 61, 62, 69, 72, 74, 79,
171, 172, 174, 179
 acquired PIK3CA mutations post
 174
 following 55, 179
Chemotherapeutic drugs 223, 469, 470
Chemotherapy 26, 250, 303, 325, 342
 arm 250
 drugs 303, 342
 regimens 26, 325
Chromosomes 4, 36, 276, 459
Cisplatin 12, 14, 18, 24, 31, 33, 45, 76,
158, 222, 224
Class-switch recombination (CSR) 327
Clones 53, 58, 73, 79, 149
 generating gefitinib-resistant 73
 resistant 53, 58, 79, 149
Coenzyme 279, 312, 411, 412
Colon cancer 34, 52, 57, 66, 69, 159,
171, 174, 184, 301, 310, 400, 401,
405, 407
 cell lines 34, 52, 57, 69, 159, 171,
174, 184
Colorectal cancer stem cells (CCSC)
291
Combination 10, 11, 13, 14, 35, 51,
173, 181, 184, 223, 224, 225, 251,
252, 290, 291, 297, 352
 cetuximab in 11, 35, 51
Combinational treatment of lapatinib
and cetuximab 184

FCDR - Anti-Cancer Agents, Vol. 3 493

Combination 217, 220, 223, 224, 225,
252, 253, 293, 294, 296, 464, 474
 of gefitinib and metformin in
 patients 293
 of Metformin 293, 294, 296
 regimens 217, 220, 223, 224, 252,
 253, 474
 therapy 223, 224, 225, 464
Complete remission (CR) 224, 238,
248, 333, 334, 340, 341, 343, 346,
354, 355
Cyclin-dependent kinase (CDKs) 252,
253, 396
Cyclophosphamide 228, 231, 235, 238,
293, 337, 343, 349
Cytochrome 277, 285, 456, 474, 475
Cytokines 28, 76, 220, 221, 227, 230,
253, 275, 329, 351, 355, 358, 360
 chemotherapeutic agents 220
 chemotherapeutic drugs 221, 253

D

Dabrafenib 219, 249, 251, 252, 296
 combination treatment of 251
Dacarbazine 217, 220, 221, 222, 223,
224, 225, 234, 237, 241, 242, 248,
249, 250, 253, 296
 arms 248, 249
 monotherapy 221, 222, 223
Data safety and monitoring board
(DSMB) 299, 303
Deacetylation 402, 403, 404
Dendritic cell (DCs) 231, 236, 237,
328, 330, 348, 351, 356
 follicular 328, 330, 351
Dendritic-cell vaccines 326
18F-Deoxyglucose positron emission
tomography 271, 272, 284, 298,
313
Dichloroacetate 289, 290, 299, 313
Dioxygenases 417, 418, 420

DNA methylation 386, 387, 389, 390, 399, 400, 401, 412, 418, 419, 423
 DNA vaccines 348, 355
 DNMT inhibitors 390, 423
 Docetaxel 38, 45, 56, 180, 222, 288, 300
 Durable tumor regressions 356, 463, 464
 Dysregulation 218, 249, 251, 395, 398, 399, 404, 409, 455

E

Eastern cooperative oncology group (ECOG) 43, 233
 Efficacy and safety of lonidamine 288, 289
 EGF expression 24, 26
 EGFR 7, 8, 15, 16, 14, 17, 18, 29, 30, 31, 32, 33, 34, 35, 38, 39, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 51, 52, 53, 54, 60, 63, 66, 68, 71, 72, 73, 80, 146, 147, 148, 149, 150, 151, 162, 163, 166, 167, 168, 169, 172, 175, 176, 177, 179, 184
 activity 60, 148, 150, 151
 blockade 63, 68, 71, 72, 184
 blockade by gefitinib in NSCLC tumors 73
 expression 8, 29, 30, 31, 32, 33, 34, 35, 38, 146, 147, 150, 151
 predictive value of 30, 31, 32
 expression and response to cetuximab 32
 expression in patient tumor tissue 29
 expression levels 29, 30, 31, 33, 34
 in cancer patient management 7
 in cetuximab-acquired resistance 149
 inhibitors 16, 17, 63, 66, 68, 162, 163, 166, 167, 168, 169, 172, 175, 176, 177
 efficacy of 150
 kinase domain mutation 17
 mutational status in NSCLC tumor tissue 41
 mutations 14, 17, 18, 39, 40, 41, 42, 43, 44, 45, 46, 47, 51, 52, 53
 activating 44, 66, 72
 common 46
 gefitinib sensitizing 179
 sensitizing 15, 16, 39, 42, 72
 tumors harbour 18
 mutations in exons 42, 43, 44
 mutations in patients 51
 point-mutation 47, 49
 sensitizing mutations 16, 18, 49, 50, 51, 80
 T790M mutations 39, 48, 49
 resistance conferring 151
 tyrosine kinase inhibitors 18, 42, 47, 48, 49, 51, 54
 EGFRvIII and PTEN co-expression 55
 EGFRvIII expression 54, 55, 56
 Endocytosis 146, 147, 148
 Endometrial Cancer (EC) 290, 293, 295, 301, 302
 Endritic cells (FDCs), (follicular 328, 330, 351
 Epidermal Growth Factor (EGF) 3, 4, 6, 24, 25, 26, 28, 29, 81, 142, 143, 145, 188, 246, 247
 Epigenetic 388, 387, 398, 399, 416, 423, 426, 427, 428
 deregulation 426, 427, 428
 dysregulation 387, 416, 423, 427
 and metabolism 388, 398, 399, 426, 427
 Epigenome 386, 387, 410, 415, 416, 418, 427
 EREG gene expression 21, 22
 Erlotinib 15, 16, 17, 18, 21, 24, 32, 33, 37, 44, 45, 51, 55, 61, 67, 70, 73, 75, 77, 78, 153, 157, 169, 172, 174, 176, 184, 185, 290

- and Metformin in Patients 290
 - efficacy 33, 73
 - Erlotinib 16, 17, 153, 169, 176
 - group 15, 45
 - monotherapy 15, 21, 75
 - resistance 67, 70
 - treatment 24, 51, 61, 70, 75, 172, 174, 184, 185
 - Evaluating IGF-1R expression 71
 - Expression levels, patient EGFR protein 30
 - Extension phase arms 248
- F**
- Fatty acids 223, 278, 279, 281, 308, 410, 411
 - FGF/FGFR signaling 72, 73
 - FGFR3 expression in gefitinib-treated cells 73
 - Fibroblast Growth Factor (FGF) 64, 72, 395
 - First-line treatment 11, 18, 51, 304
 - First-line treatment of patients 11, 12
 - FL-cells 325, 327, 329, 330, 331, 338, 353
 - survival 325, 330, 331
 - FL 325, 326, 328, 330, 332, 342, 343, 344, 347, 355, 358
 - clones 328, 330, 358
 - relapse 342, 343, 344
 - treatment 325, 326, 328, 332, 347, 355
 - FOLFIRI 11, 15, 61, 154, 163, 291
 - FOLFOX4 155, 163
 - Fuel molecules 277, 279, 281
 - Fumarate hydratase (FH) 285, 286, 298, 313, 387, 397, 421, 422, 423
- G**
- Gain-of-function mutations 5, 392
 - Gastric cancer 25, 244, 299, 391, 399, 400
 - Gefitinib 14, 20, 30, 31, 36, 38, 40, 44, 45, 49, 50, 55, 58, 60, 61, 67, 69, 70, 73, 147, 148, 177
 - efficacy of 31, 69, 73, 147, 177
 - evaluating 14, 36, 38, 44, 55, 147
 - inhibitory effects of 40, 60, 70
 - Gefitinib 9, 18, 20, 28, 30, 31, 38, 40, 41, 43, 44, 45, 46, 48, 49, 54, 56, 61, 67, 69, 70, 71, 73, 77, 148, 165, 166, 172, 176, 179, 184
 - and erlotinib 9, 18, 40, 54, 56, 61, 67, 70, 77
 - inhibition 41, 176
 - resistance 71, 165, 184
 - conferred 148
 - response 20, 30, 31, 38, 70
 - sensitivity 179, 184
 - treated cells 73
 - treatment 28, 43, 44, 45, 46, 48, 69, 71, 166, 172
 - first-line 44, 172
 - relapsed NSCLC tumor post 49
 - Gemcitabine 14, 18, 33, 38, 45, 172, 290, 294, 297, 470
 - Gene copy number (GCN) 35, 39, 82, 174, 188
 - Gene expression 21, 59, 285, 386, 387, 389, 390, 392, 393, 397, 403, 404, 411, 413, 423
 - Gene therapy 459
 - Genes 20, 21, 22, 36, 218, 276, 386, 387, 389, 390, 391, 398, 404, 459, 472
 - fusion 276
 - metabolic 285, 286, 427
 - tumor suppressor 246, 286, 387, 390, 391, 402, 414
 - Glioblastoma 42, 54, 176, 177, 286, 291, 297, 299, 309, 405
 - Glioma patients 74, 405, 408, 418
 - Glutamine 394, 395, 398, 406

Glutaminolysis 387, 394, 395, 398, 403, 406, 409, 412, 417
 Glycolysis 278, 279, 280, 281, 306, 307, 394, 397, 398, 399, 400, 404, 405, 406, 412
 GM-CSF group 231
 Grade Lymphoma Study Group (GLSG) 340
 Groups 22, 23, 35, 46, 147, 148, 225, 228, 232, 235, 236, 239, 240, 243, 246, 252, 354, 389, 392, 413, 419
 dabrafenib-trametinib 252
 methyl 389, 392, 413, 419
 vaccine 235, 236
 vaccine-IL-2 232

H

Harbouring K-RAS mutations 158
 HB-EGF expression 26, 27
 HDAC inhibitors (HDACi) 391, 424
 Hematologic malignancies 294, 295, 299, 304, 351, 357, 397, 475, 476, 477
 advanced 294, 295, 299, 304
 HER2 13, 29, 51, 57, 58, 59, 158, 171, 173, 181
 amplification 57, 58, 59
 expression 29, 58, 173
 FISH 58
 -positive breast cancer patients 173, 181
 -positive metastatic breast cancer patients 13, 51, 171, 173
 HER3 expression 61, 62, 73
 Hexokinase 279, 284, 299, 302, 307, 313, 395, 397, 405
 High EGFR expression 30, 32
 Himeric antigen receptors (CARs) 350, 351

Histone 386, 387, 390, 391, 392, 402, 404, 410, 411, 412, 413, 415, 419, 420, 421, 424
 acetylation 387, 391, 410, 411, 412, 424
 demethylases 386, 387, 419, 420
 methylation 387, 392, 404, 412, 413, 415, 419, 420, 421
 modifications 386, 387, 390, 392, 402, 404, 421
 Histones 390, 391, 392, 412, 415, 426
 HKII expression 405
 H-RAS mutations 152, 159
 Human cancers 227, 275, 282, 388, 389, 390, 392, 393, 399, 404, 406, 409, 421, 424, 427
 Human growth factor (HGF) 28, 64, 65, 66, 395
 2-R-hydroxyglutarate 417, 418, 419, 426, 427
 Hypoxia-inducible factor (HIF) 285, 388, 395, 396, 397, 399, 401, 404

I

IAP antagonists 467, 470, 472
 IAP proteins 466
 IDH1 286, 295, 299, 417, 418, 419, 420
 mutations in 417, 419
 wild type 418, 420, 426
 IDH1 and IDH2 286, 304, 312, 417, 425
 mutations 286, 312
 IDH1 mutations 299, 304, 417, 420
 IDH1 mutations and AG-221 in patients 304
 IDH2 mutations 286, 299, 304, 312, 417
 IDH2-R140Q, expressing 426
 IDH-mutant glioma cells 423, 424
 Idiotype vaccines 354

Subject Index

IGF-1R expression 70, 71
IGF2 expression 69
IGFBP3 expression 70
III trials, randomised Phase 14, 18, 44
II trials, randomized phase 224, 231, 341
IL-2 227, 228, 229, 231
 administration of 229
 high-dose 227, 228, 231
IL-2 group 232
Immune 10, 150, 217, 226, 227, 231, 232, 238, 239, 240, 244, 245, 246, 252, 274, 329, 330, 331, 347, 348, 351, 354, 355, 356, 357, 359, 427
 cells 226, 274, 330, 347, 348, 427
 checkpoint 217, 226, 238, 239, 240, 244, 245, 246, 252
 response 10, 150, 227, 231, 232, 240, 329, 330, 331, 351, 354, 355, 356, 357, 359
Immunohistochemistry 20, 21, 28, 29, 30, 31, 32, 33, 34, 35, 64, 65, 177, 178, 179
Immunotherapeutics 220, 225, 231, 242
Immunotherapy 217, 219, 220, 275, 325, 326, 338, 347, 348, 350, 351, 357, 358, 359, 459, 460
 active 347, 348, 359
 passive 326, 347, 348, 350
Induction chemotherapy 290, 295
Induction regimens 333, 338, 339, 340, 341, 343, 345
Inhibitors 10, 51, 52, 217, 226, 239, 240, 244, 249, 250, 253, 273, 276, 396, 398, 399, 420, 422, 424, 458
 immune checkpoint 217, 226, 239, 240, 244
Insulin-like growth factor (IGF) 28, 64, 68, 249
Interleukin-2 217, 226, 227, 234
Ipilimumab 217, 219, 240, 241, 242, 243, 244, 245, 246, 252, 253, 275

FCDR - Anti-Cancer Agents, Vol. 3 497

 receiving 241
Ipilimumab group 240, 241, 243, 245
Ipilimumab monotherapy 245
Irinotecan 10, 11, 13, 15, 21, 27, 33, 34, 62, 69, 154, 156, 291, 294, 469
Isocitrate 286, 416, 417
Isocitrate dehydrogenase 279, 286, 298, 299, 307, 313, 387, 416

J

JmjC histone demethylases 419, 420, 421

K

Ketogenic diets 272, 288, 291, 294, 295, 307, 308, 309
Ketone bodies 278, 279, 307, 309
Khambata-Ford 21, 158, 166
Kinase domain, tyrosine 5, 40, 42, 43, 143
K-RAS mCRC patients 171
KRAS mutations 155, 158
K-RAS mutations 11, 21, 57, 154, 155, 156, 157, 158, 159, 160, 181, 182, 186
 association of 154, 181
 common 155
 harboured 154, 156
 mCRC harbouring 155
K-RAS mutations 152, 155, 158
 in mCRC 152, 155
 sub-population of patients 158
K-RAS wildtype mCRC patient tumors 182

L

Lactate dehydrogenase 279, 281, 298, 313, 397, 400

- Lactate production 311, 400, 402, 405, 406, 424
- Lactic acid fermentation 271, 277, 278, 279, 280, 281, 283, 284, 311
- LAK cells 227, 228, 326, 348, 349, 352
- Lapatinib 9, 13, 23, 24, 59, 60, 64, 78, 79, 171, 172, 173, 180, 184, 185
and capecitabine 13, 24
- Lenalidomide 331, 345, 358
- Leukemias 251, 332, 406, 464, 465, 474, 476
- Ligands 3, 4, 6, 7, 19, 20, 22, 23, 28, 29, 80, 81, 143, 144, 146, 242
and receptors 6, 19, 80, 81, 146
- Localisation, nuclear 146, 147, 149, 150
- Lonidamine 288, 289, 299, 306
- Lung cancers, non-small cell 183, 291, 299, 302, 303, 308, 313, 406
- Lymphokine-activated killer (LAK) 227, 228, 325, 348, 352
- Lymphomas 272, 337, 349, 352, 358, 471, 474, 476
- Lysate, vaccinia melanoma cell 236
- Lysine 50, 391, 392
- M**
- mAbs 7, 9, 10, 64, 146, 157, 188
- Macrophages, tumor-associated 329, 330
- Maximum tolerated dose (MTD) 248, 249, 476
- MCRC patients 12, 13, 21, 22, 24, 25, 26, 27, 28, 33, 35, 52, 53, 58, 60, 65, 66, 71, 75, 76, 154, 156, 157, 161, 168, 170, 178, 179, 182
chemorefractory 71
chemo-refractory 12
chemotherapy-refractory 156
cohort of 27, 35, 65, 75, 178, 179, 182
expressing 13
irinotecan-refractory 168
naïve 53
randomised chemo-refractory 154
refractory 21
stratified 21
third-line treated 76
treated chemo-refractory 27
untreated 66
- MDM2 inhibitors 409, 460, 463, 465
- Median progression 222, 224, 225
- Median survival 218, 222, 224, 229, 334
- Median TTP 11, 14, 15, 58
- MEK Inhibitors 250, 253
- Melanoma 217, 218, 219, 220, 225, 228, 229, 230, 241, 244, 246, 247, 248, 251, 400
- Melanoma development 218
- Metabolic 281, 286, 287, 299, 301, 302, 307, 311, 382, 386, 394, 396, 399, 404, 407, 409, 410, 412, 413, 425, 427
enzymes 281, 286, 287, 386, 412, 425, 427
pathway 299, 301, 302, 311, 394, 399, 404, 409, 410, 413
plasticity 282, 307
stress 396, 407
- Metabolism 271, 272, 277, 283, 284, 285, 307, 310, 388, 393, 395, 398, 399, 402, 403, 404, 406, 409, 412, 424, 425, 426, 427, 428
altered 395, 404, 409, 425, 427
central carbon 271, 272, 277, 283, 284, 285
- Metabolites 409, 416, 422, 426, 427
- Metastatic melanoma 218, 220, 222, 223, 225, 226, 227, 228, 229, 235, 237, 238, 240, 241, 253
treatment of 217, 220, 221, 222, 229, 230

Subject Index

Metastatic pancreatic cancer 18, 290, 292, 297, 302
Metformin 293, 301, 304, 305, 306
Metformin 272, 289, 290, 291, 292, 293, 294, 295, 296, 297, 301, 304
on colorectal cancer stem cells 291
Methylation 387, 390, 391, 401, 402, 410, 413, 415
Methyl donor 412, 413, 414
Mitochondrial succinate dehydrogenase 387
Mitogen activated protein kinase (MAPK) 164, 165, 166, 188, 246
Molecular mechanisms 3, 80, 142, 146, 186, 305, 455
Monoclonal antibodies 7, 9, 188, 275, 325, 326, 331, 350, 351, 352, 357
Monotherapy 13, 15, 37, 44, 217, 223, 346, 347, 478
Multicenter phase II Trial 290, 344
Mutant IDH1 417, 418, 420, 425
Mutational analysis 42, 143, 187
Mutational status 28, 172, 182, 183, 186
Mutations 21, 40, 42, 43, 45, 46, 48, 49, 50, 52, 53, 151, 155, 160, 164, 167, 170, 171, 175, 182, 395, 397, 402, 422
acquired 48
activating 40, 42, 43, 52, 59, 151, 160, 164, 167, 168, 171, 174, 175, 186, 246, 253, 285, 304, 309, 395, 388, 389, 391, 392, 397, 402, 421, 422, 423
common 46, 155
driver 388, 389, 392
genetic 174, 246, 253, 402, 422
identified 40, 167, 168
inactivating 309, 391, 397, 421
lethal cancers harbor 304
novel 52, 186
oncogenic 285, 420
somatic 40, 59, 175, 246, 423

FCDR - Anti-Cancer Agents, Vol. 3 499

tumors harbouring KRAS 21
Mutations 386, 421
in FH 421
in metabolic enzymes in cancer 386
MYC expression 397, 398, 408

N

National comprehensive cancer network (NCCN) 155, 158, 333, 337
Negative metastatic breast cancer patients 293
Nicotinamide N-methyl-transferase (NNMT) 414, 415
Nivolumab 219, 242, 245, 246, 253
NK cells 231, 325, 326, 348, 349, 350, 352
Non-coding RNA 386, 387, 392
Non small cell lung cancer (NSCLC) 14, 15, 16, 41, 42, 43, 49, 50, 51, 61, 153, 244, 297, 299, 302
North-east Japan study group (NEJSG) 14
Novartis 465, 470, 471
N-RAS mutations 152, 156, 159, 160, 182
NRG2 β 4, 6, 9, 64, 145
NSCLC cell lines 20, 62, 70, 72, 73, 77, 79, 147, 151, 172, 180
NSCLC patient harbouring EGFR mutations 18, 51
NSCLC patients harbouring EGFR sensitizing mutations 16, 65
Nutlin-3 461, 463, 465

O

Oncometabolites 286, 386, 388, 416, 417, 424
ORR and TTP of mCRC patients 179
ORR in cetuximab patients 155

Ovarian cancer 75, 244, 308, 310, 400
 Oxaliplatin and Cetuximab in
 First-Line Treatment of mCRC 11
 Oxidative phosphorylation 278, 279,
 307, 313, 388, 394, 395, 396, 398,
 410

P

Panitumumab efficacy 12, 65, 66, 172
 Paraganglioma 286, 421, 423
 Patient cohort 26, 179, 340
 Patient-derived xenograft (PDX) 57,
 66, 69, 79, 82, 165, 172, 188
 Patient-matched tumor-normal pairs 42
 Patient populations 71, 80
 Patients 14, 33, 36, 38, 44, 52 55, 67,
 76, 156, 157, 161, 163, 170, 178,
 228, 229, 231, 235, 242, 244, 304,
 305
 asymptomatic 331, 332
 bronchioloalveolar carcinoma 33, 36
 cetuximab-naïve 52
 chemotherapy-naïve 33
 chemotherapy-naïve 14, 33, 157
 diabetic 304, 305
 vaccinated 231, 235
 Patients harbouring K-RAS mutations
 158
 Patient's post-cetuximab treatment 185
 Patient tumors 151, 173
 untreated NSCLC 66
 Patient tumor tissue analysis 186
 Patient tumor tissue post-treatment 78
 PD-1 239, 241, 242, 357
 PDGFR β 73, 74
 expression 73
 gene expression levels in
 post-cetuximab 74
 PDX tumors, cetuximab-sensitive 79
 Pembrolizumab groups 243

Pentose phosphate pathway (PPP) 287,
 398
 Pheochromocytoma 285, 421, 422, 423
 Phosphorylated IGF-1R 69, 70, 71
 PIK3CA 57, 167, 168, 169, 170, 171,
 172, 173, 174, 175, 181, 182, 183,
 186
 exon 170, 171, 174, 182
 gene 167, 168, 170, 174, 406
 mutational status 168, 172, 173
 mutation and response to cetuximab
 168
 mutations 57, 167, 168, 169, 170,
 171, 172, 173, 174, 175, 181, 183,
 186
 acquired 174
 harboured 167, 170
 role of 172, 173
 PIK3CA 172, 173, 174
 mutations and pathological
 complete response 173
 mutations and pathological
 complete response in HER2 173
 mutations in breast cancer cell lines
 172
 mutations in NSCLC patient
 biopsies 174
 Platinum based chemotherapy 295, 296
 Post-cetuximab 58, 74, 160, 185
 Predictive biomarker 20, 21, 55, 59, 62,
 66, 80, 150, 163, 164, 177, 182,
 186
 Pro-apoptotic protein 454, 455, 458
 Progenitor cells 47, 420, 421
 Progression-free survival 82, 188, 223,
 246, 248, 249, 252, 337
 Progression free survival (PFS) 22, 45,
 76, 158, 161, 163, 168, 170, 224,
 225, 245, 339, 341, 343, 346
 Progressive disease 23, 65, 69, 75, 76,
 238, 291
 Promoter DNA methylation 389, 390,
 400, 402

Subject Index

Promoter methylation 389, 399, 400, 401, 402
Protein expression 22, 29, 34, 64, 75, 148, 472
Protein phosphatase 407
Proteins 144, 147, 246, 248, 275, 355, 358, 389, 390, 392, 402, 458, 459, 464, 465, 466, 472, 474
 histone 390, 402
 tumor suppressor 389, 458, 465
PTEN 55, 144, 145, 167, 171, 172, 174, 175, 176, 177, 178, 179, 180, 181, 182, 395, 400, 407, 408
 function of 171, 181
 expression 55, 172, 177, 178, 179, 180, 400, 407
 expression and cetuximab efficacy 178
 expression and patient response 180
 expression status 178, 181
 loss 171, 174, 175, 176, 177, 178, 179, 400
Pyruvate 279, 281, 284, 287, 299, 307, 313, 397, 398, 400, 410, 463
 dehydrogenase 279, 281, 307, 313, 397, 410
 kinase (PK) 279, 284, 287, 299, 307, 313, 398, 400, 463

R

Radioimmunotherapy 325, 326, 333, 337, 338, 340, 341, 342
RAS mutations 156, 157
Renal cell cancer (RCC) 228, 401, 404, 421
Renal cell carcinoma 241, 244, 401, 404, 421
Resistance 19, 28, 47, 50, 61, 67, 69, 70, 72, 74, 78, 79, 146, 165, 166, 172, 175, 183, 186, 474
 development of 70, 78, 474

FCDR - Anti-Cancer Agents, Vol. 3 501

 mediated 50, 69, 70, 79
Resistance mechanisms 3, 5, 57, 61, 63, 66, 72, 142, 146, 164, 179, 182, 183
 evaluated 61, 66
 pre-existing intrinsic 142, 146
Resistance mediators 72, 152, 157
Resistance mutations 39, 51, 152
 acquired 51
 common acquired 39
 rare acquired 39
Resistant cells 28, 70, 74, 79, 148, 149, 159, 165, 185
Response to cetuximab 24, 26, 27, 33, 34, 35, 69, 158, 168, 178, 179
Response to gefitinib 14, 30, 36, 55, 58, 63, 66, 177
Restriction, caloric 288, 307, 308, 309
Review, systematic 37, 161, 163, 170, 171
R-2-hydroxyglutarate 420, 421, 422
Rituximab 335, 336, 337, 338, 339, 340, 341, 342, 343, 344, 345, 346, 350, 352, 353

S

S-adenosyl-homocysteine (SAH) 413, 415, 426, 427
S-adenosylmethionine 387, 389, 392, 410, 412
SCCHN cell lines 21, 23, 27, 34, 58, 61, 165, 166, 172, 185
SCCHN tumors 42, 43, 59
Scorpion amplification refractory mutation system (SARMS) 48
Sensitizing mutations 46, 47
Serum of NSCLC patients 65, 180
Silencing, epigenetic 399, 401, 402
SMAC mimetic 464, 468, 469
Squamous cell lung carcinoma (SCLC) 8, 37, 62, 474

Stage III melanoma 226, 233, 236
Stage IV melanoma 219, 220, 242, 243,
244, 249, 250, 251, 252
STAT3 activity in gastric cancer cells
185
Sub-clones, stable cetuximab-resistant
H226 149
Sub-populations 14, 23, 25, 28, 57, 80,
187
cetuximab-resistant A431 73
Succinate 286, 397, 416, 421, 422, 423
Succinate and fumarate 416, 421, 422
Survivin 466

T

Targeted cancer therapies 272, 274,
277
Targeting apoptosis pathways 454, 455
Taxanes 13, 158, 220, 222, 224, 273,
290
T cell-receptor (TCRs) 329, 350
T-cells 226, 325, 350
anti-tumor 234, 238
Temozolomide 221, 222, 223, 224,
234, 291, 303
TET methyl-cytosine dioxygenase 387
TGF α 6, 9, 21, 22, 23, 24, 64, 82, 188
Therapeutic strategies 325, 329, 331,
356, 359, 425, 455
Therapy 21, 26, 28, 46, 56, 65, 71, 76,
142, 145, 154, 161, 163, 165, 168,
169, 171, 172, 175, 177, 178, 179,
183, 186, 187, 218, 219, 228, 238,
239, 240, 246, 253, 272, 275, 299,
301, 303, 312, 331, 335, 338, 339,
348, 350, 352
adoptive cell 238, 331, 348, 352
based 46, 65, 71, 154, 161, 162,
163, 169, 171, 172, 177, 186, 187,
335
cetuximab-based 76, 168, 178, 179
metabolic cancer 272, 299, 303
targeted 21, 26, 28, 56, 165, 175,
177, 183, 186, 218, 219, 246, 253,
272, 275
Threonine 48, 50, 415
Toxicities 15, 223, 229, 236, 242, 252,
276, 338, 348, 455
Trametinib 219, 250, 251, 252
Transcription factors 81, 246, 247, 281,
284, 285, 388, 389, 396, 397
Translocation 327, 328, 390, 392, 419
Tricarboxylic acid cycle 387
Tri-methylation 392, 415, 420, 422
Tumoral cells 349, 353, 357
Tumor antigens 233, 234, 235, 348,
350, 353
Tumor-associated antigens (TAAs)
230, 234, 353, 359
Tumor-associated macrophages (TAM)
77, 329, 330, 479
Tumor-infiltrating lymphocytes (TILs)
228, 238, 359
Tumor necrosis factor (TNF) 227, 228,
237, 457, 469
Tumor progression 23, 47, 48, 80, 331
Tumor regression 18, 74, 229, 230,
272, 465, 471
Tumor resistance 19, 29, 47, 56, 68, 73,
81, 142, 146, 150, 187, 273
acquired 47, 142
Tumors 15, 21, 22, 23, 32, 33, 37, 42,
49, 50, 52, 53, 57, 58, 60, 63, 69,
74, 78, 79, 80, 148, 157, 159, 173,
177, 178, 179, 234, 242, 245, 246,
272, 283, 287, 302, 348, 349, 354,
419, 423, 464, 469, 476, 477
cetuximab-refractory 60
erlotinib-resistant 78
expressing 32, 157, 173
mutation-positive 245
negative 37, 58, 63, 69, 242, 246
patients harbouring 173
primary 42, 49, 159, 178, 179

Subject Index

recurrent 49, 148
refractory 74
resistant 50, 78
solid 15, 302, 348, 349, 423, 464,
469, 476, 477
treated 52, 53
Tumor suppressors 285, 390, 398, 399,
401, 403, 458, 460
Tumor tissue 40, 55, 59, 71, 75, 167,
176, 184, 185, 282, 421
colorectal carcinoma patient 167
evaluated patient 176

U

UNC Lineberger comprehensive cancer
center 293, 295

V

Vaccines 230, 233, 237, 353, 355, 356
dendritic cell 237, 356
idiotypic 353, 355
peptide 230, 233
Vaccinia melanoma cell lysate
(VMCL) 236
Vaccinia melanoma oncolysate (VMO)
236
Vascular endothelial growth factor
(VEGF) 28, 64, 74, 75, 76, 225,
395, 461
VEGF expression 74, 76
VEGF gene expression 75
VEGF levels 76
Vincristine 223, 290, 291, 337
Vitro efficacy of cetuximab 23, 34

W

Warburg effect 271, 272, 277, 282,
284, 287, 288, 298, 299, 305, 306,
308, 310, 312, 397
Warburg's phenotype in cancer cells
424
Wt K-RAS 22, 57, 154, 155, 156, 157,
158, 159, 161, 168, 182
Wt K-RAS subgroup 22, 156

X

Xenografts 66, 74, 82, 188
generated cetuximab resistance
A431 74
patient-derived 66, 82, 188



PROF. DR. ATTA-UR-RAHMAN, *FRS*

Atta-ur-Rahman, Ph.D. in organic chemistry from Cambridge University (1968), has 1020 international publications in several fields of organic chemistry including 727 research publications, 37 international patents, 68 chapters in books and 188 books published largely by major U.S. and European presses. He is the Editor-in-Chief of eight European Chemistry journals. He is Editor of the world's leading encyclopedic series of volumes on natural products "Studies in Natural Product Chemistry" 48 volumes of which have been published under his Editorship by Elsevier during the last two decades.

Prof. Rahman won the UNESCO Science Prize (1999) and was elected as Fellow of the prestigious Royal Society (London) in July 2006. He has been conferred honorary doctorate degrees by many universities including (Sc.D.) by the Cambridge University (UK) (1987), He was elected Honorary Life Fellow of Kings College, Cambridge University, UK, conferred the TWAS (Italy) Prize and the Austrian government has honoured him with its high civil award ("Grosse Goldene Ehrenzeischen am Bande") (2007). He is Foreign Fellow of Chinese and Korean Academy of Sciences, Foreign Fellow of the Chinese Chemical Society and former President of Pakistan Academy of Sciences.